



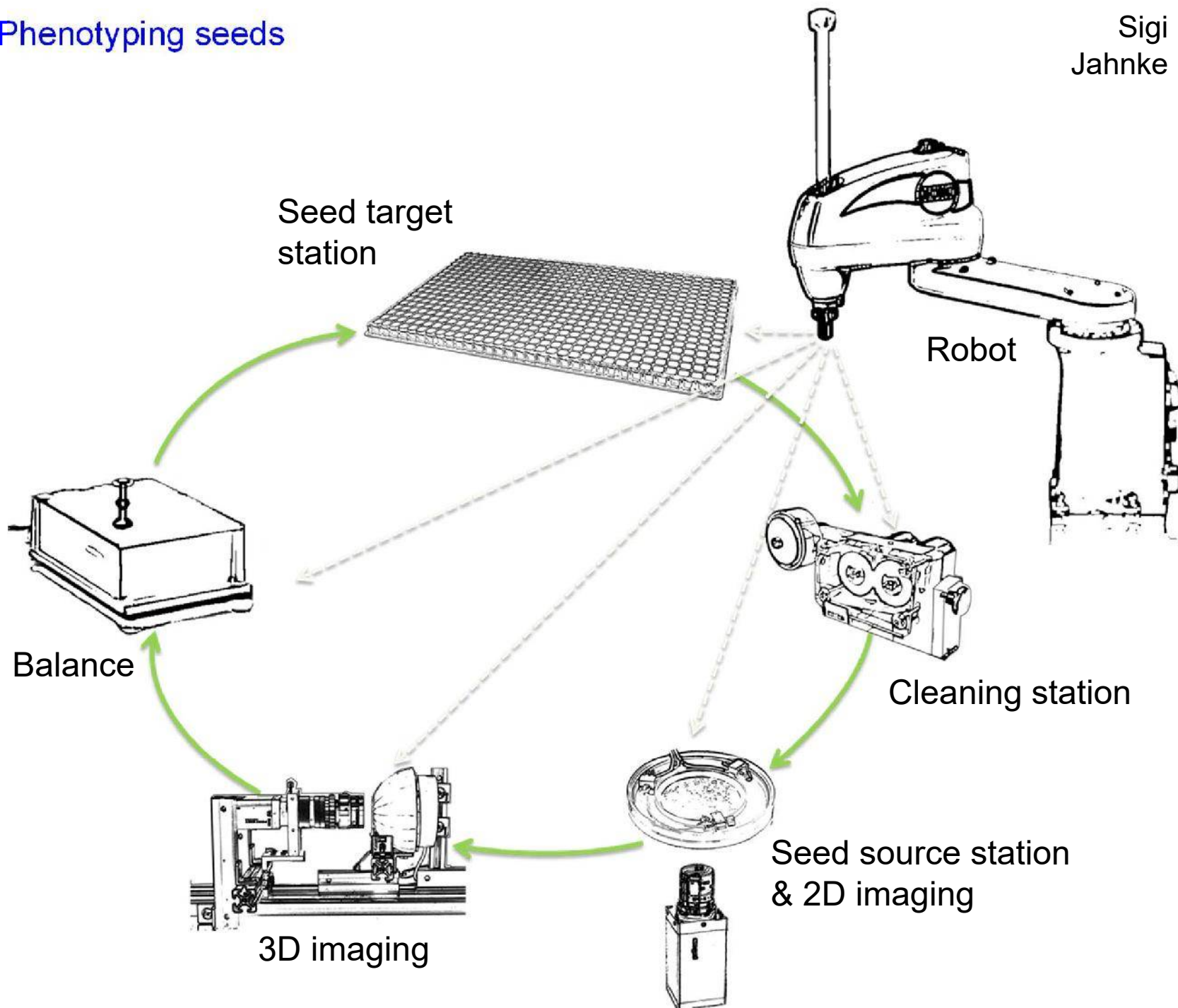
1. Genes, phenes and machines @ FZJ
2. Bridging the gap between lab and field

Poorter et al. (2016)
Pampered inside, pestered outside?
Differences and similarities
between plants growing in
controlled conditions and in the field.
New Phytol. 212: 838-855

Uli Schurr
Hendrik Poorter

1. Phenotyping seeds

Sigi Jahnke

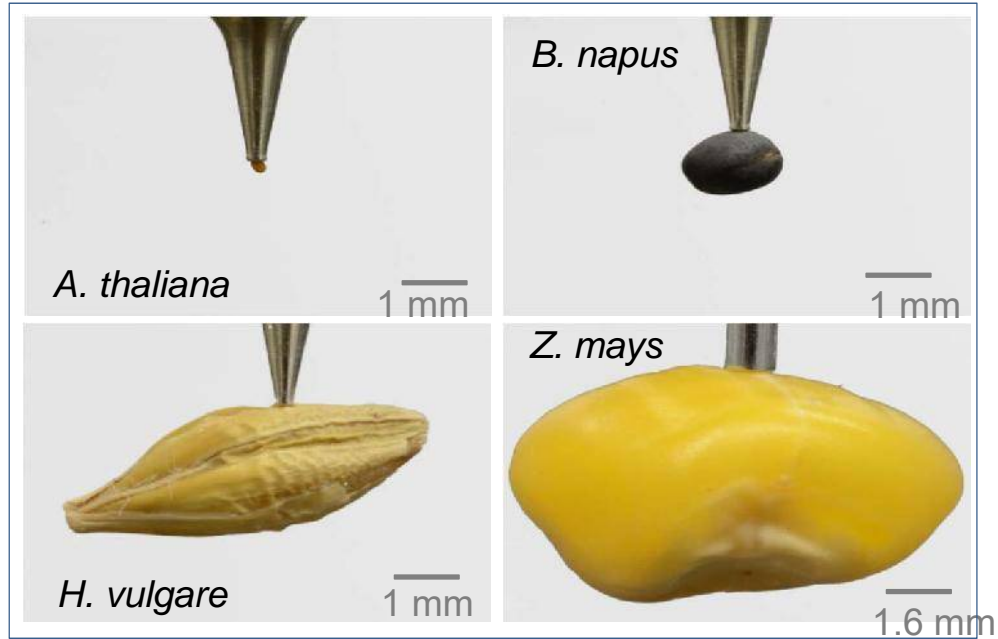


1. Phenotyping seeds



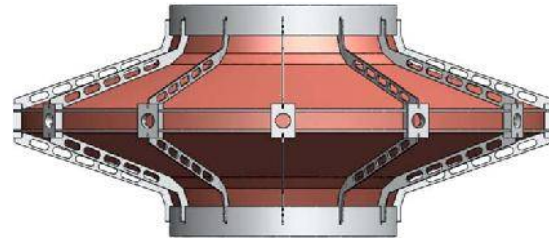
1. Phenotyping seeds

Jahnke et al. (2016)
Plant Phys., in press



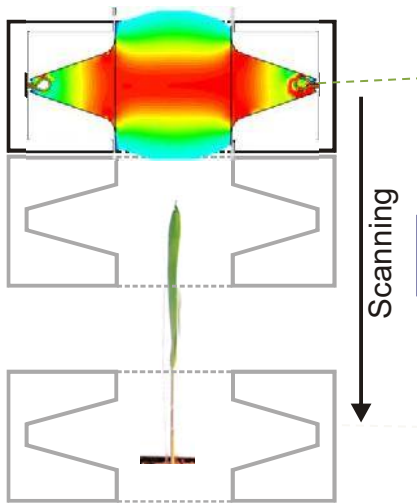
	WT	<i>etr1</i>	P
Plant mass (mg)	35	14	***
RGR (mg g ⁻¹ day ⁻¹)	260	268	ns
Ind. seed mass (µg)	20	13	

2. Measuring water



Microwave cavity resonator

Viktor
Sydoruk



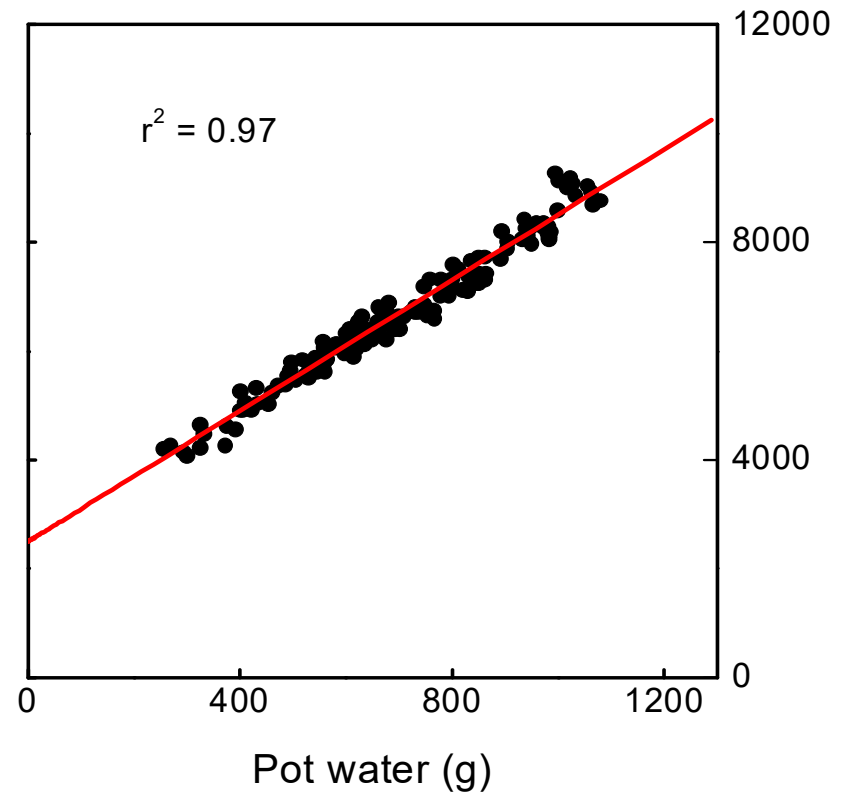
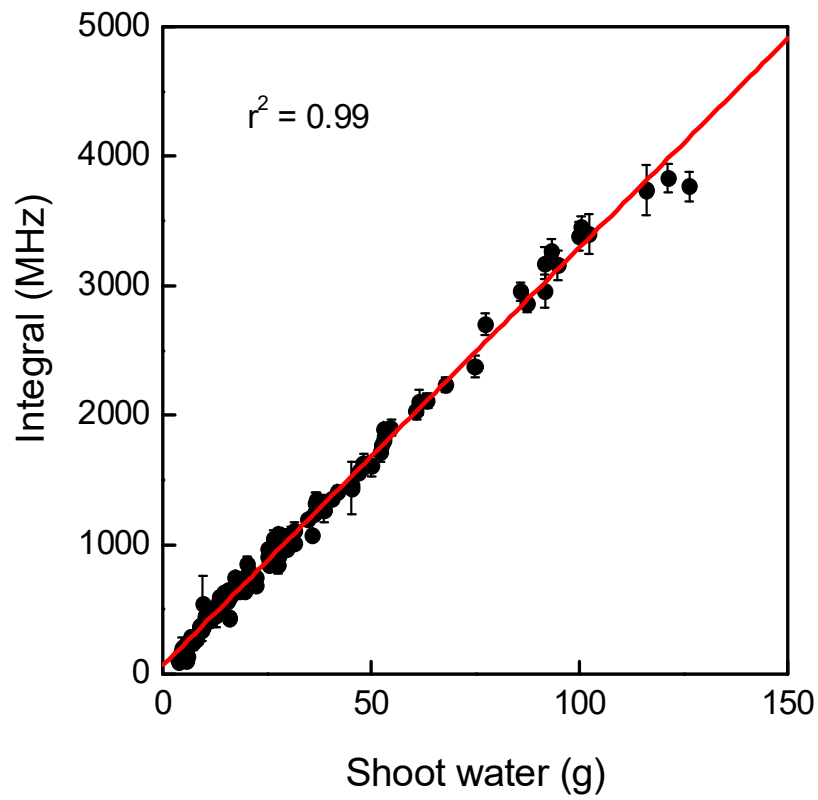
- Homogeneous field in x,y plane
- High sensitivity in x,y plane
- Narrow range of sensitivity in z direction

2. Measuring water

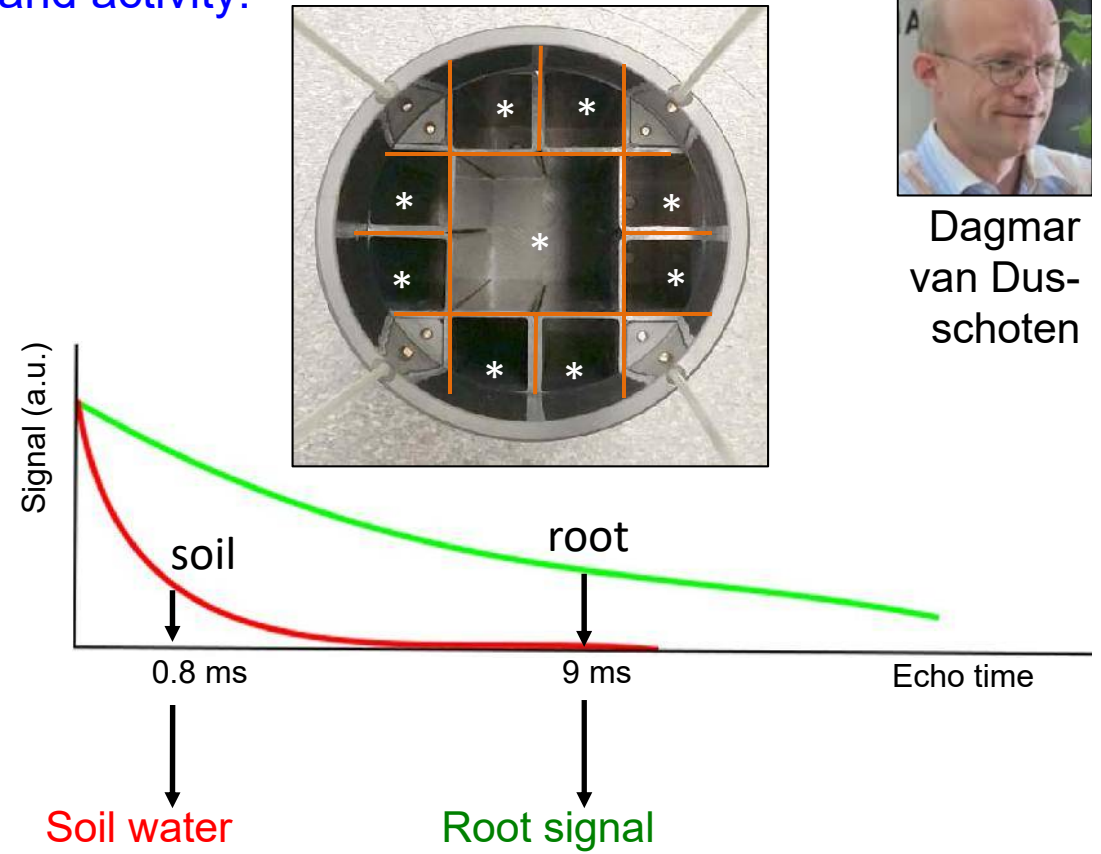
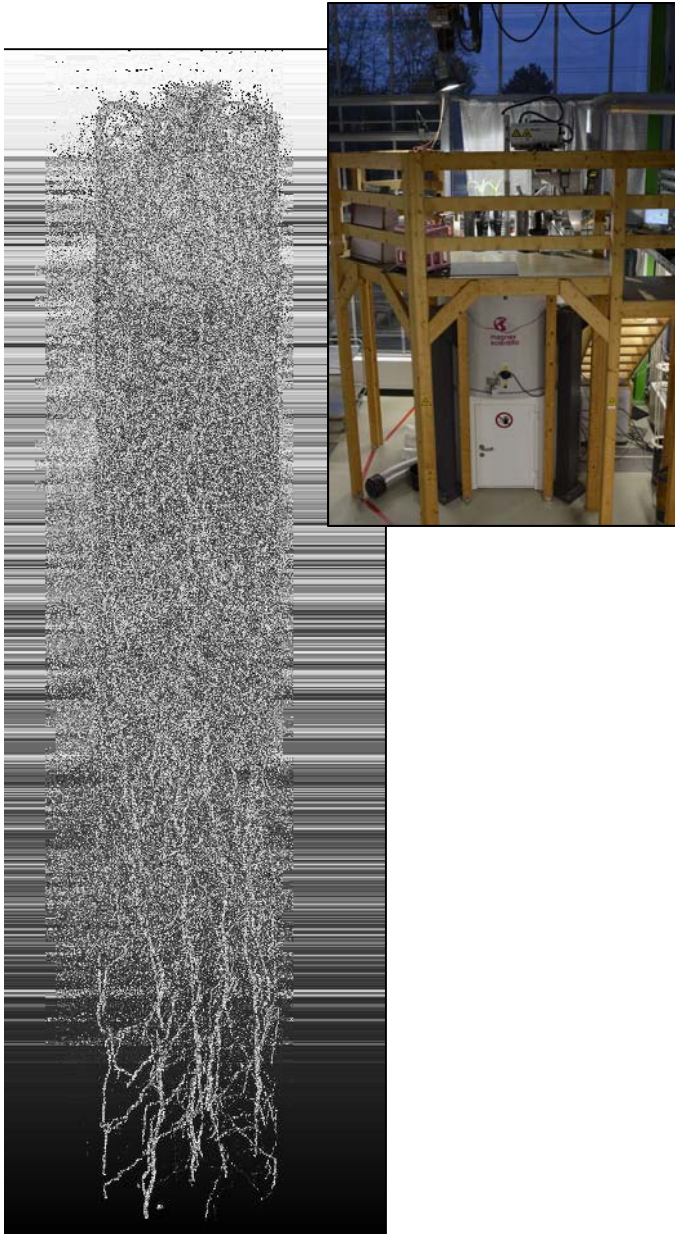


2. Measuring water

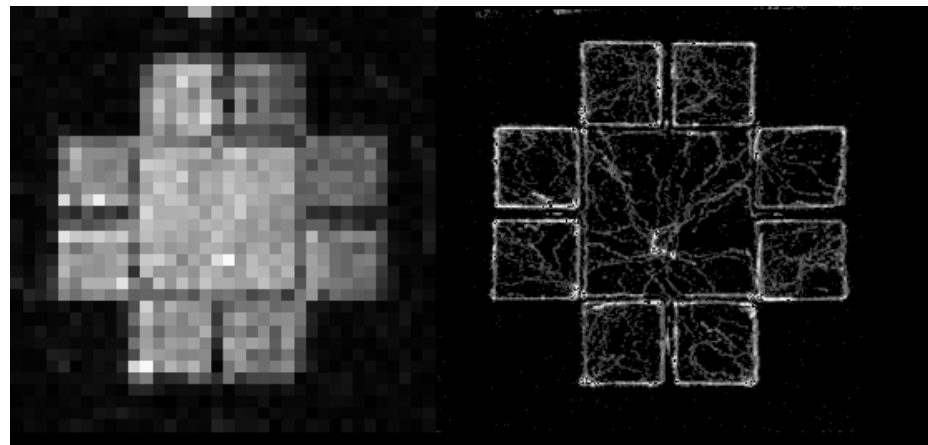
Sydoruk et al. (2016)
IEEE Transact. 64: 2894



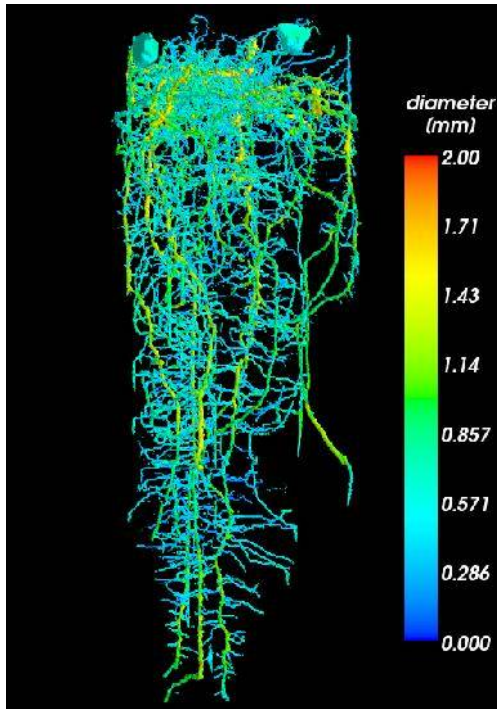
3. Measuring spatial root distribution and activity:



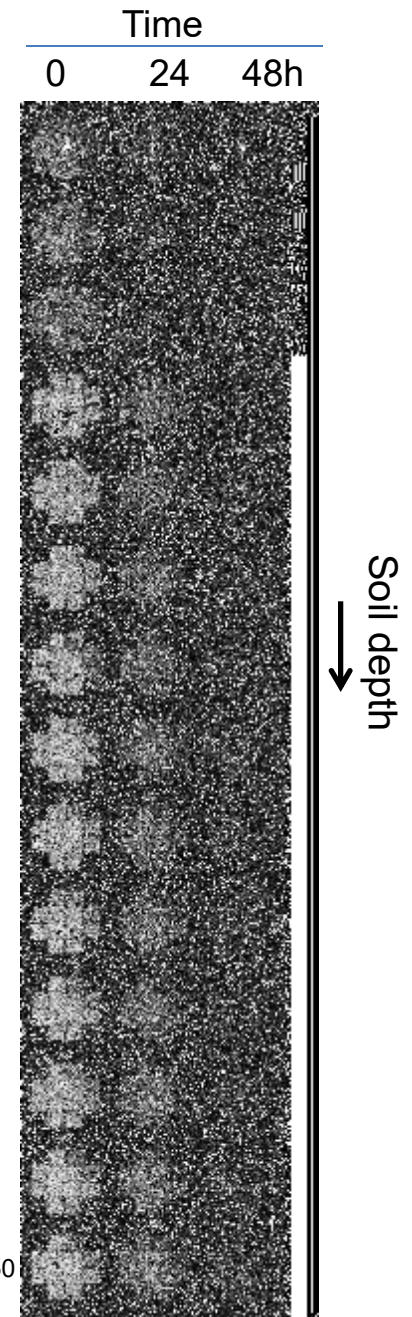
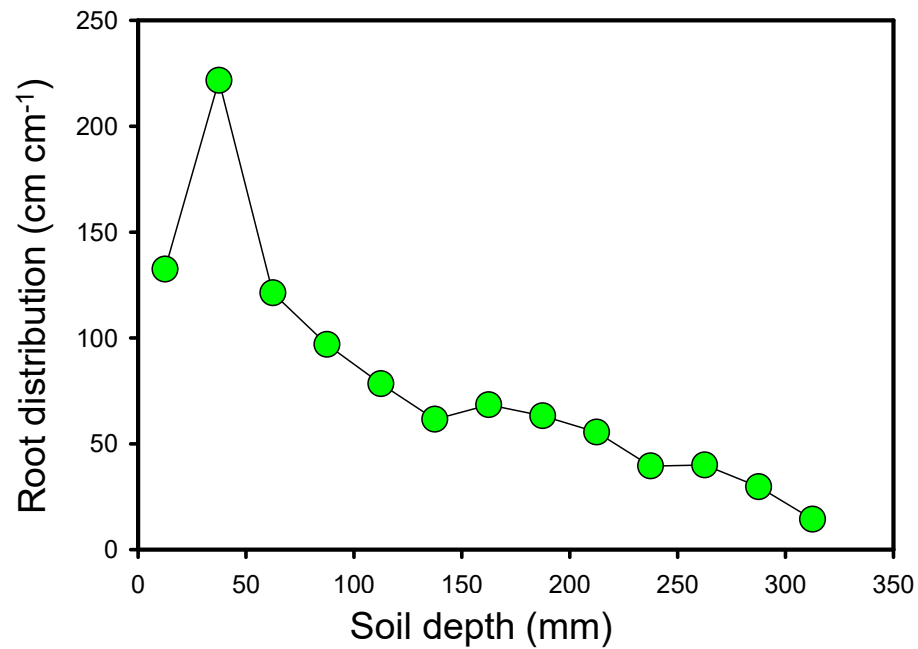
Dagmar van Duschoten



3. Measuring spatial root distribution and activity:

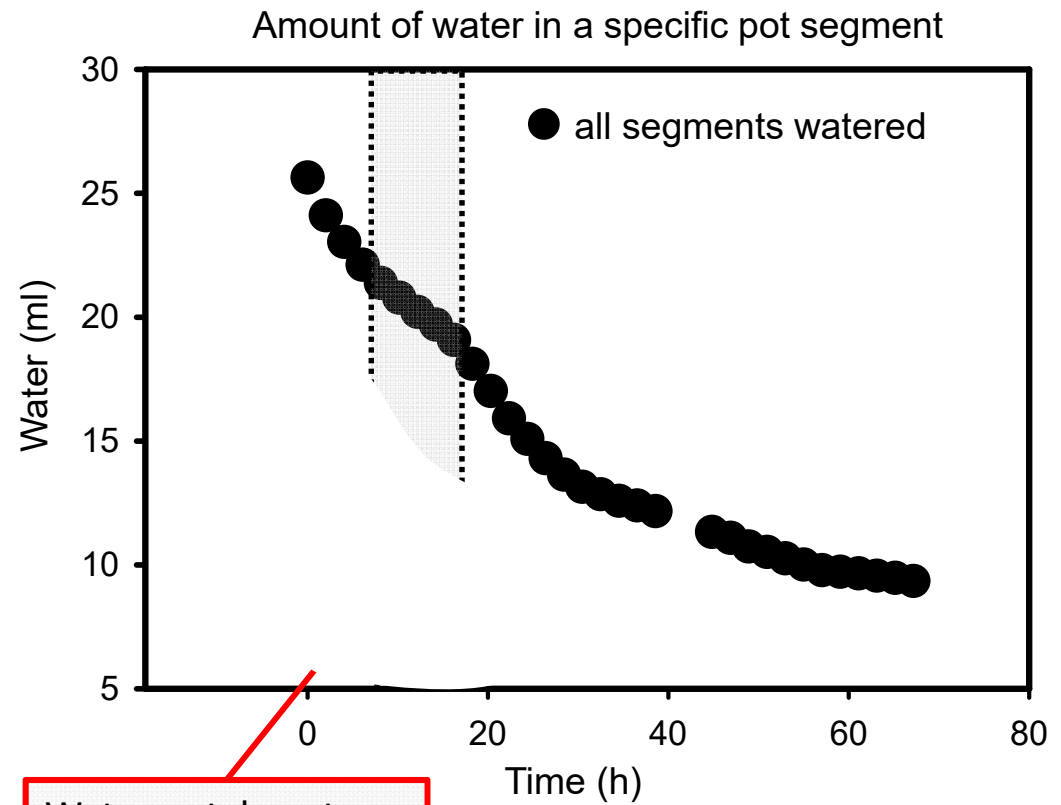
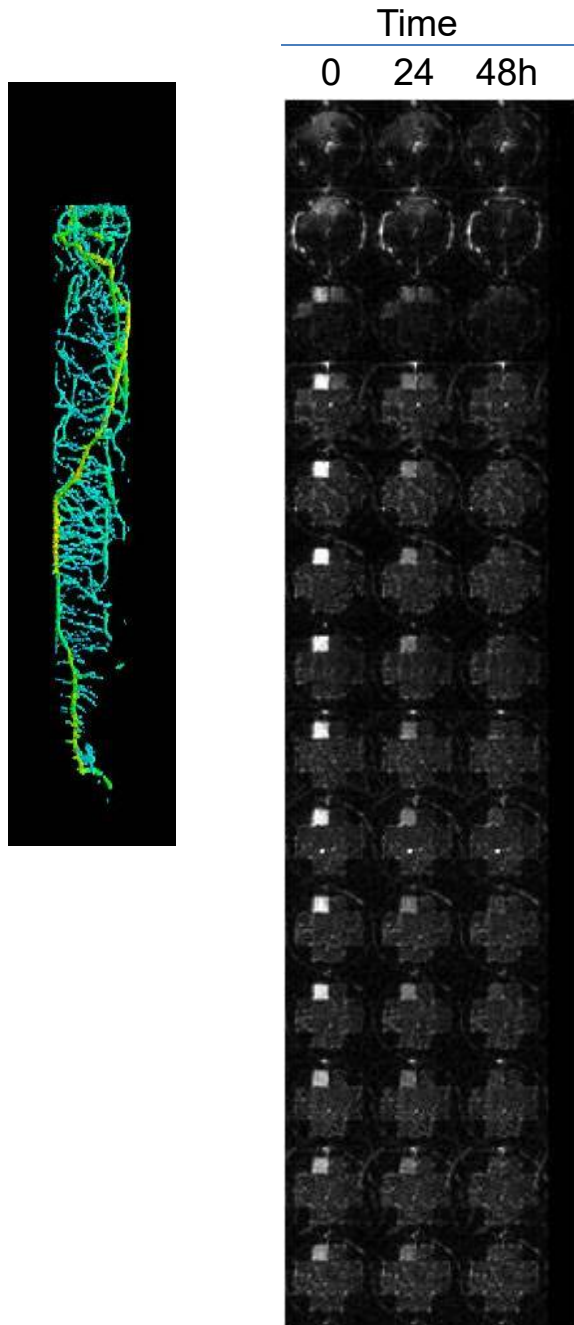


Total length: 26.7 m
Root tips: 1460
Avg. branch angle: 70-75°
Fresh mass: 6.1 gram



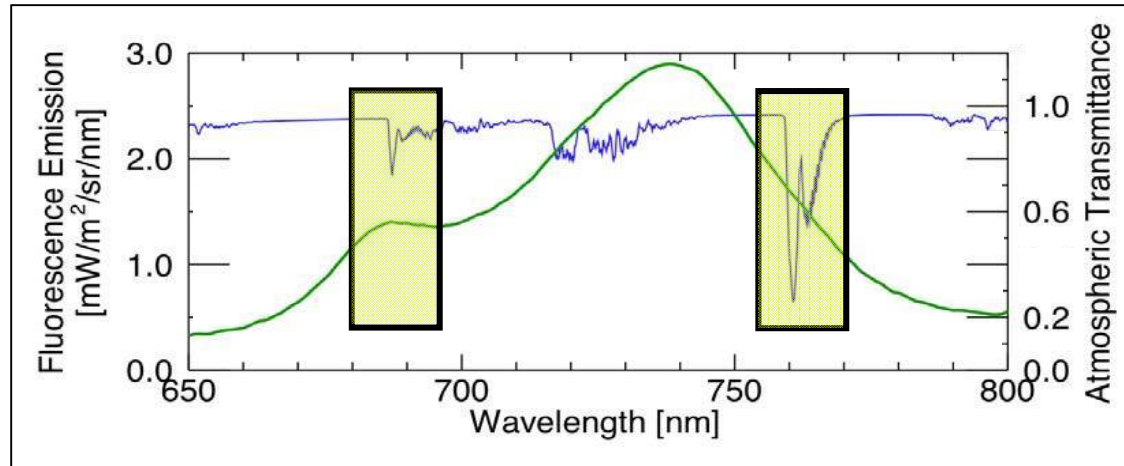
3. Measuring spatial root distribution and activity:

Van Dusschoten et al.
Plant Physiol. 170: 1176



Water uptake rate:
~7.5 times faster

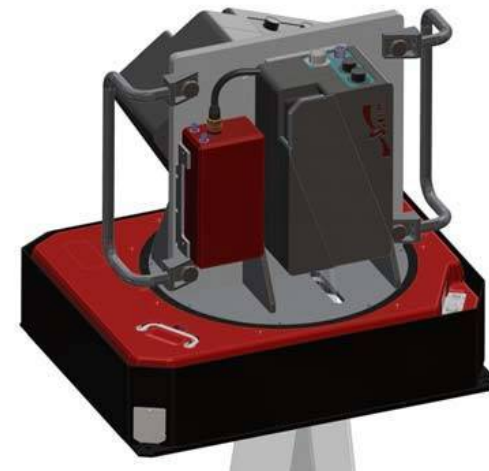
4. Sun-induced fluorescence:



Uwe
Rascher

HyPlant:

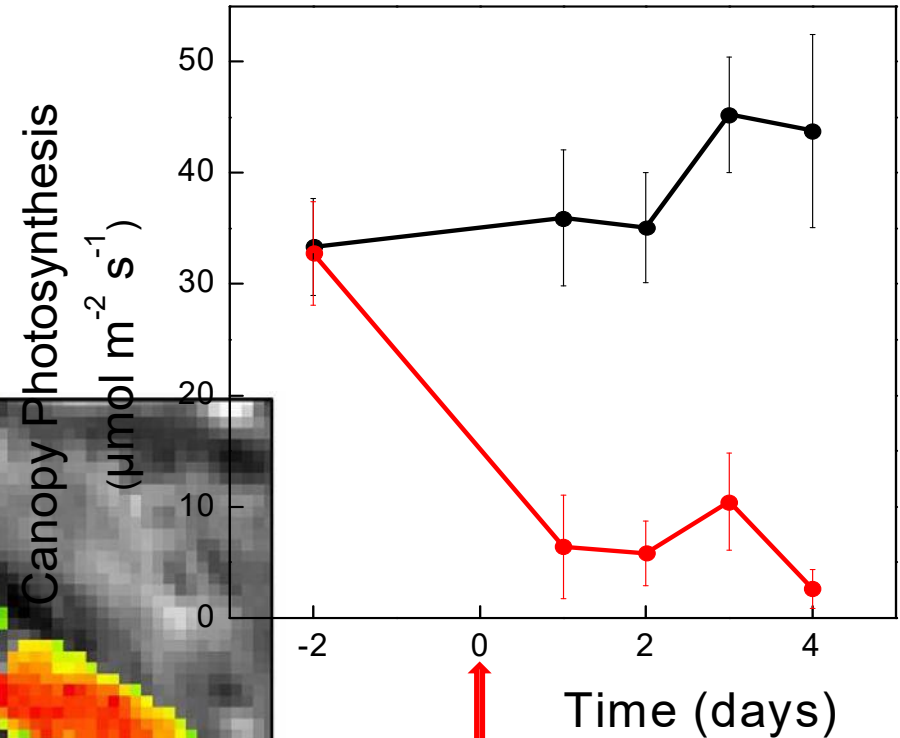
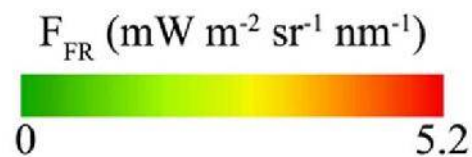
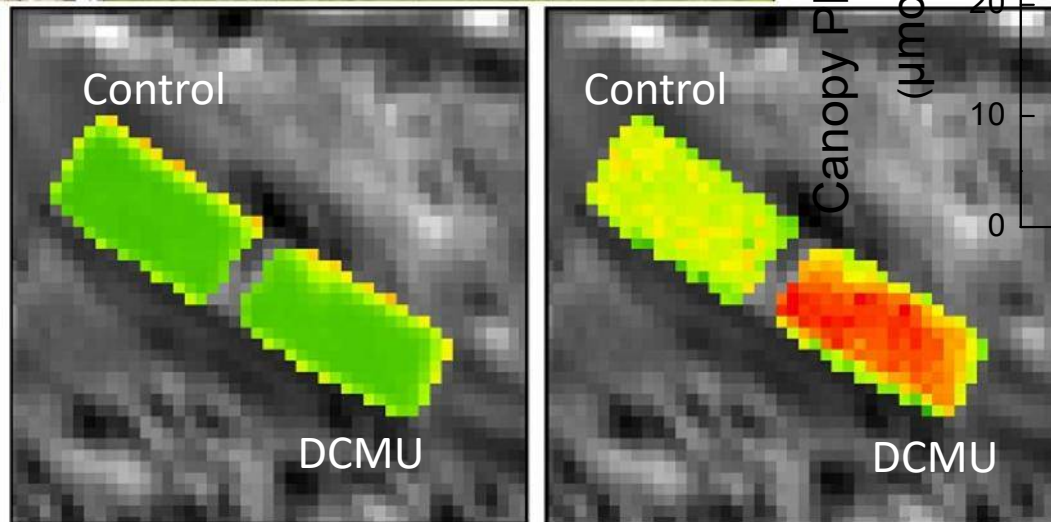
- Module 1:
Broad spectrometer (380 - 2500 nm)
- Module 2:
High-resolution fluorescence
module (670 - 780 nm)



4. Sun-induced fluorescence:

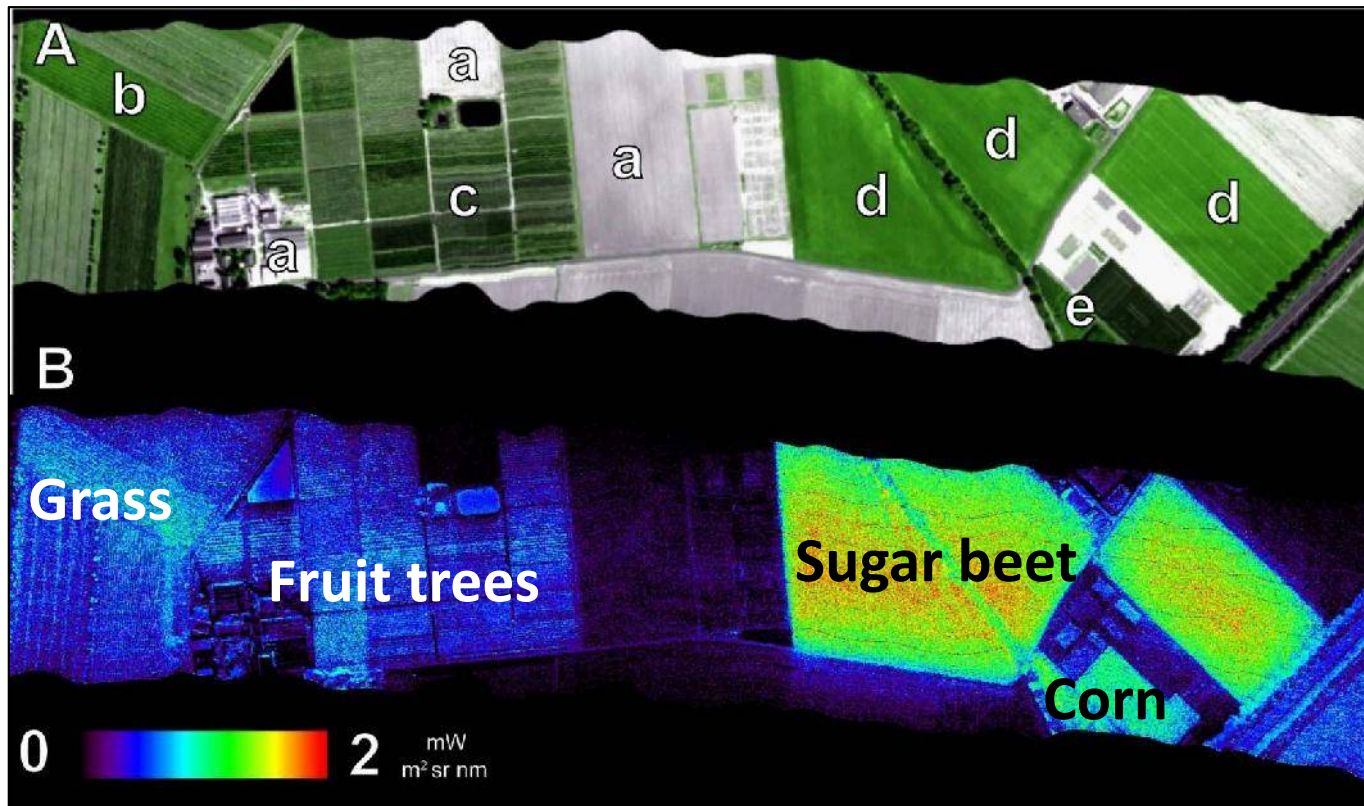
Rossini et al. (2015)
Geophys Res. Lett. 42

- 2x 140 m² grass carpets
- Add DCMU to one



+ DCMU

4. Sun-induced fluorescence:



Flex satellite mission 2022 (ESA):

- Direct measurements vegetation fluorescence
- Pixel size: 300 x 300 meter
- In tandem with vegetation temperature
- Covering each location on earth every 10-25 days

Creating a sustainable Plant Phenotyping Community in Europe:



National platforms



European projects/
networks



ESFRI

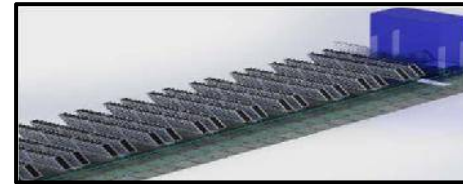
European Infrastructure

European infrastructure for Multi-site
plant PHenotyping And Simulation for
food Security in a changing climate

Emphasis infrastructure:



- Lab-based platforms for high resolution, high throughput phenomics



- Semi-controlled field systems for high throughput phenomics

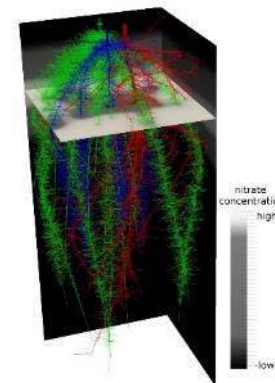


- Network of field sites for lean-phenotyping all over Europe

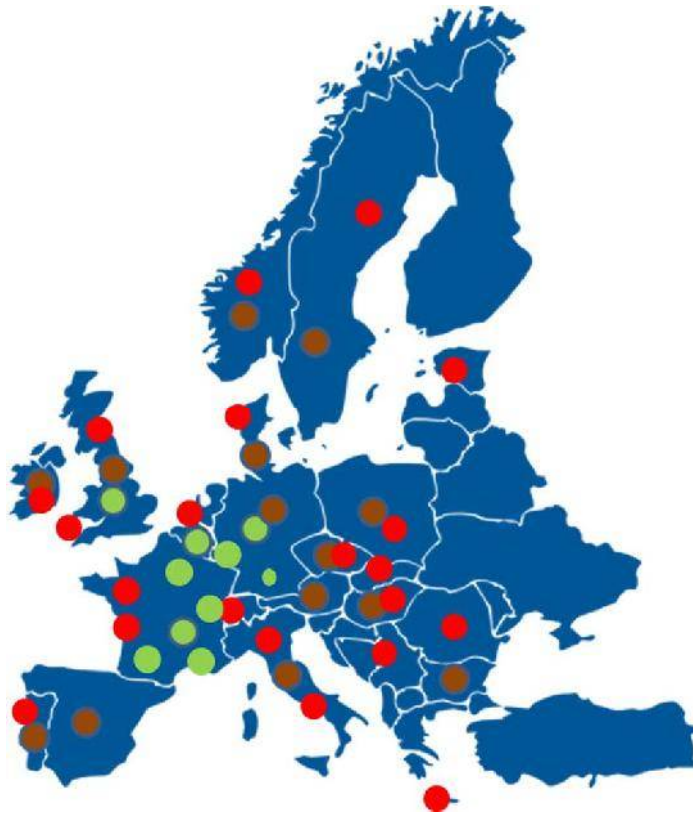


- Joint data management and e-infrastructure

- Modelling for improving phenotypic processes and for testing existing or virtual combinations of alleles in a variety of climatic scenarios and management practices



Emphasis: development



- EMPHASIS project partners
- Field sites
- EMPHASIS associated partners

with support from



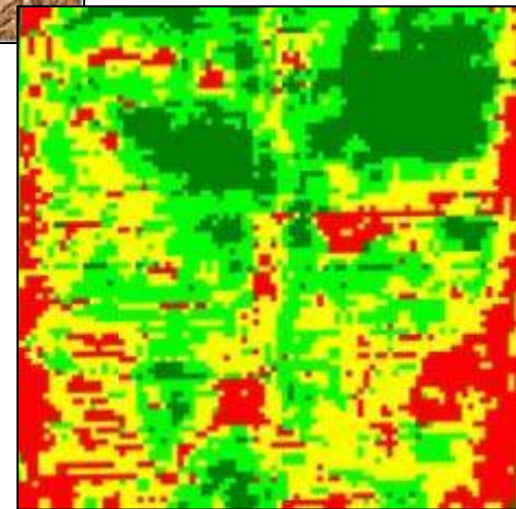
Field networks of seed companies

EMPHASIS - open for additional partners

The problems of growing plants outside:



Normalised yield

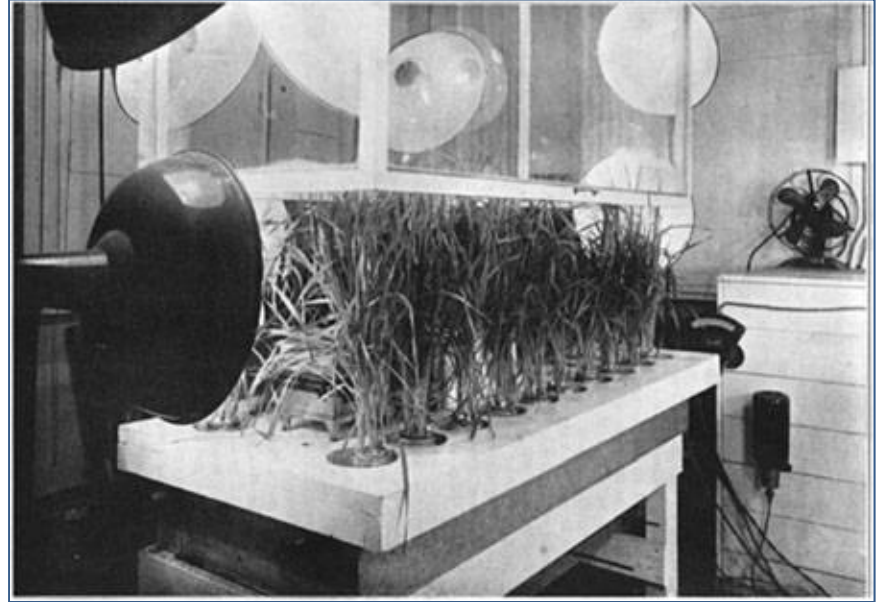


Large spatial variation

Large temporal variation

Strong improvements in environmental control:

Davis & Hoagland (1928)



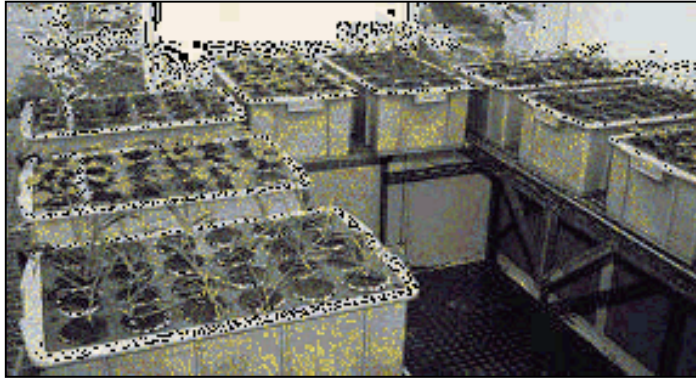
modernfarmer.com

Questions to us scientists:

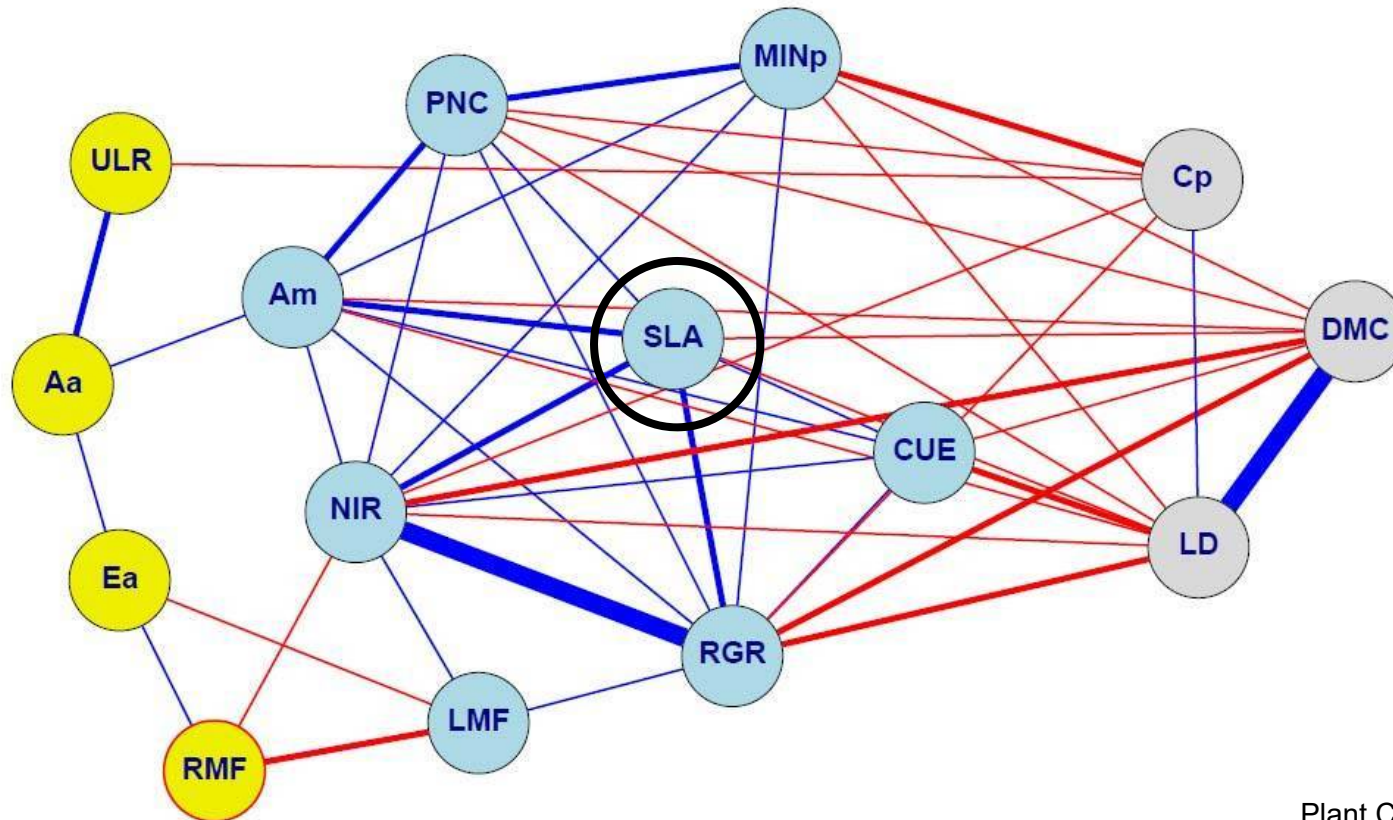


- 1. How different are plants growing in controlled conditions and in the field?**
- 2. How can the differences be explained?**
- 3. What can we do to improve the lab-field correlation?**

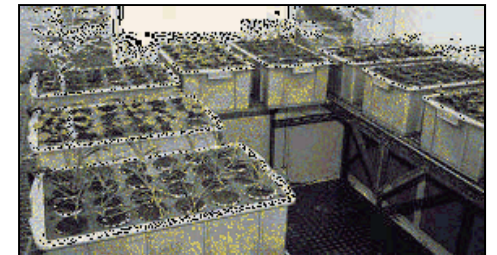
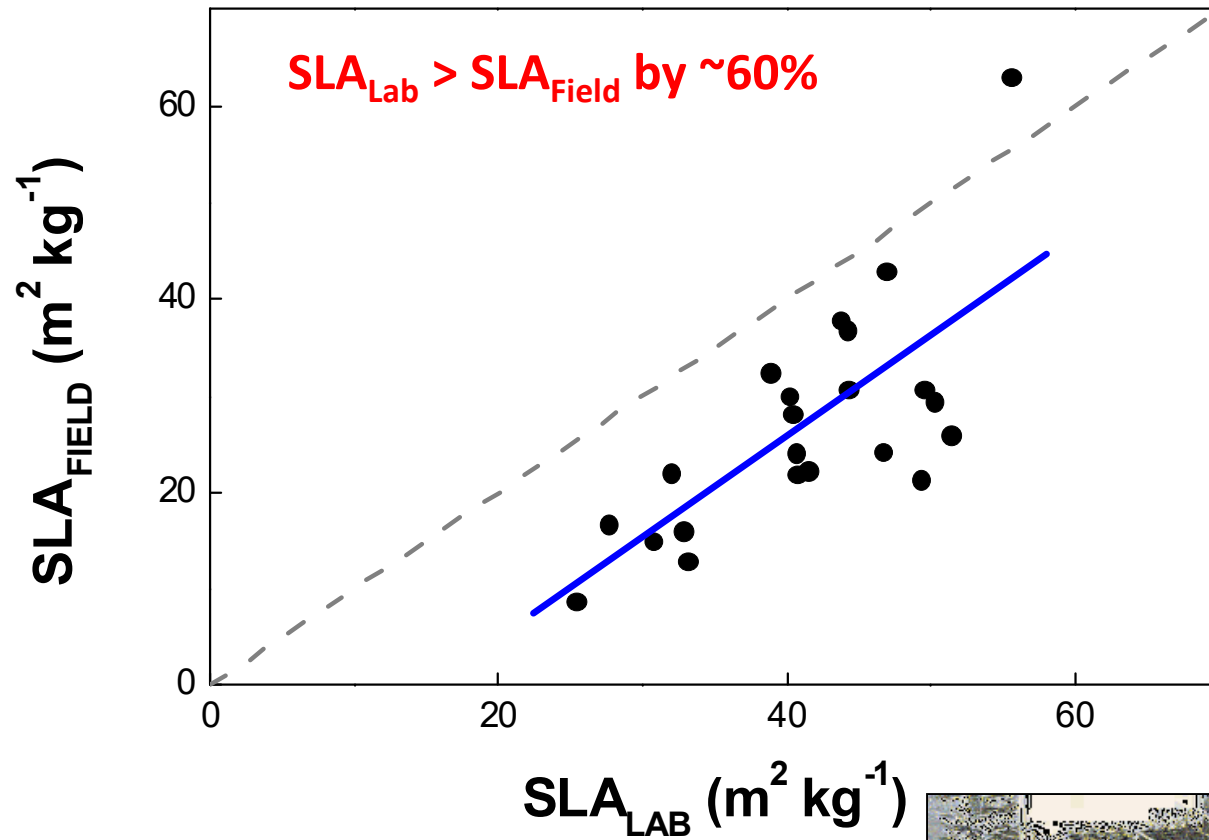
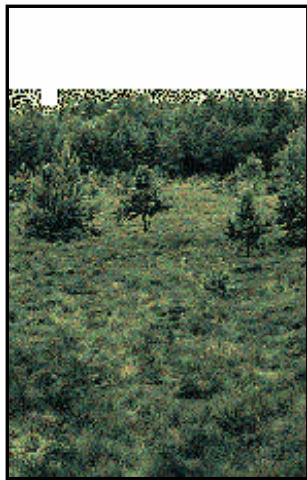
Q1: Do lab and field plants have similar phenotypic values?



- $T_{\text{Day}} = T_{\text{Night}} = 20\text{ }^{\circ}\text{C}$
- Irradiance = $315\text{ }\mu\text{mol m}^{-2}\text{ s}^{-1}$
- Daylength = 14 h
- RH = 70%
- Nutrients = 2 mM NO_3^-
- 24 species



Q1: Do lab and field plants have similar phenotypic values?



Q1: A 'brute force' approach:

Meta-Phenomics database:



- > 1000 controlled experiments
- > 1100 species
- ~ 10 traits
- 12 environmental factors

LEDA database:



- field data
- > 1500 species
- ~ 27 traits

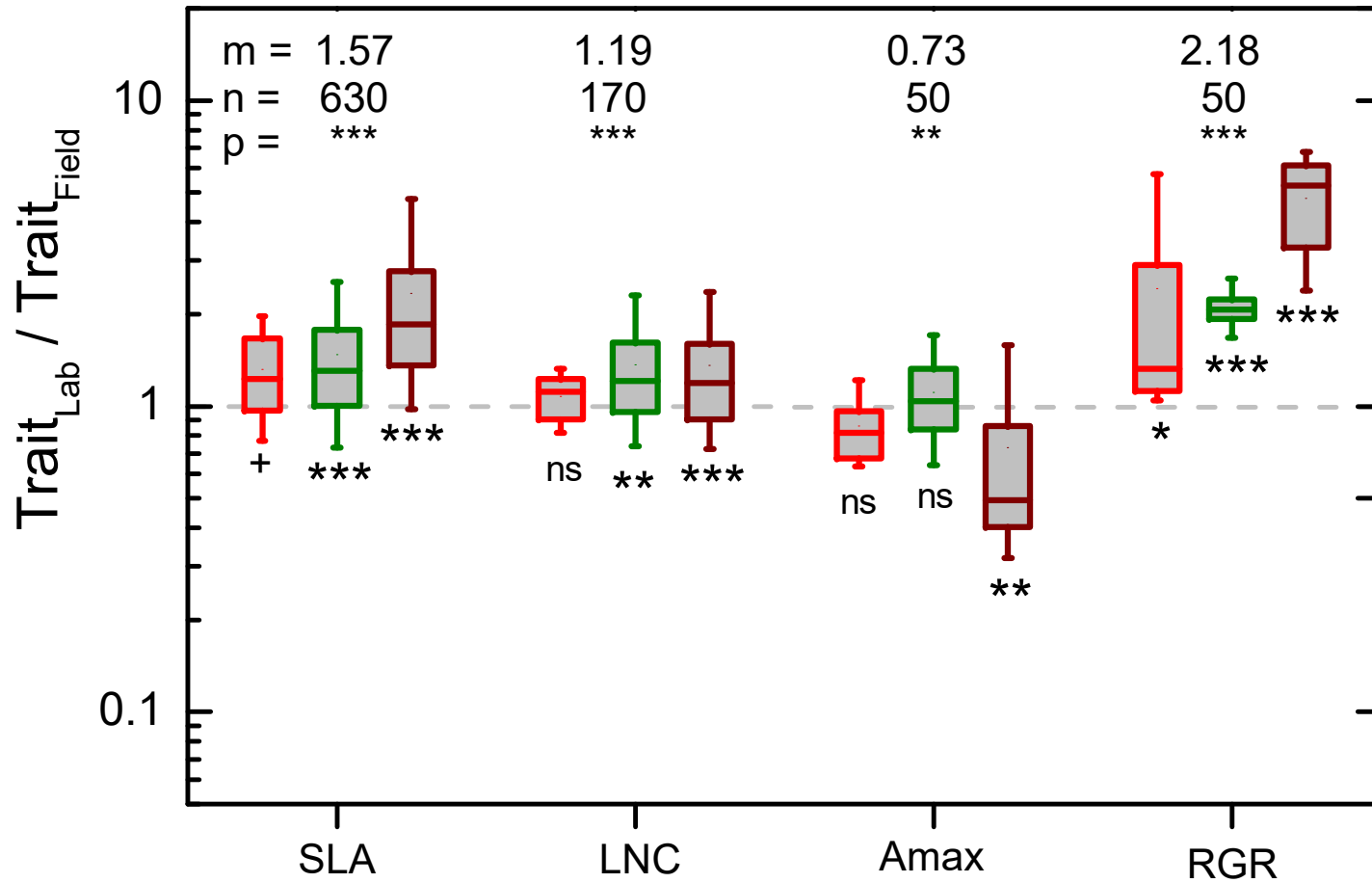
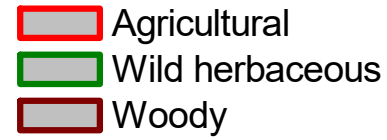
Other compilations & sources:

- Niinemets (2001)
- Wright et al. (2004)
- Poorter et al. (2009)
- Many additional papers (crop species!)

Targeted database:

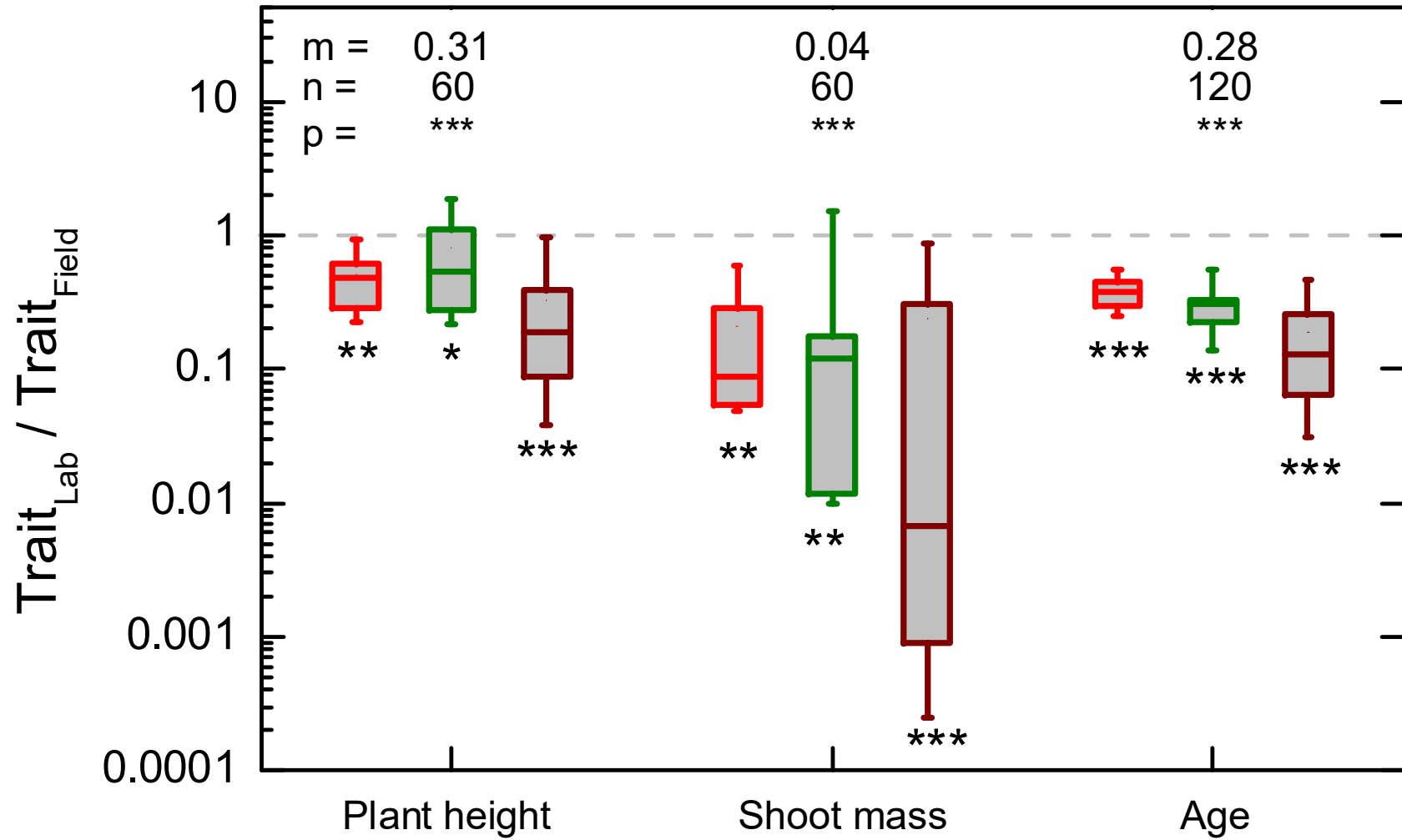
- 7 traits
- 20000 records
- > 5500 species
- > 1500 references

A meta-analysis across species:



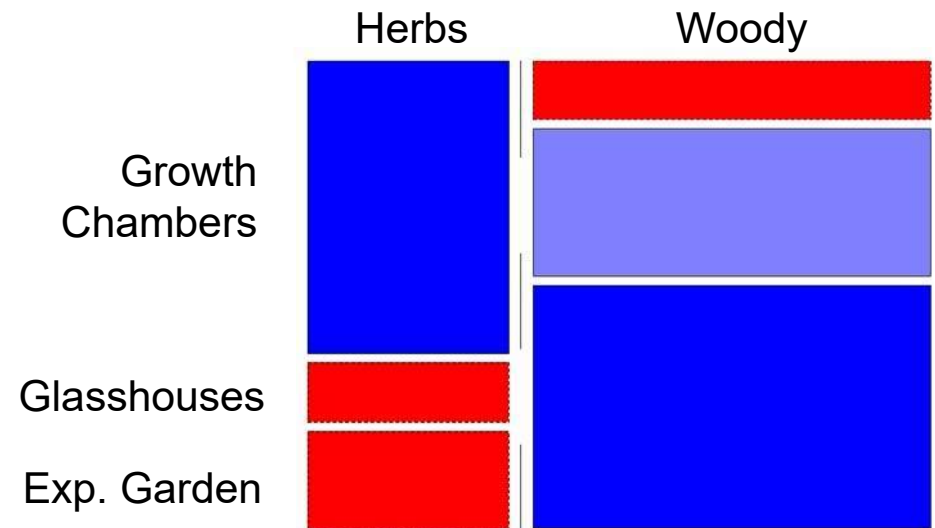
Size-related parameters:

Median life span in GC:
39 days

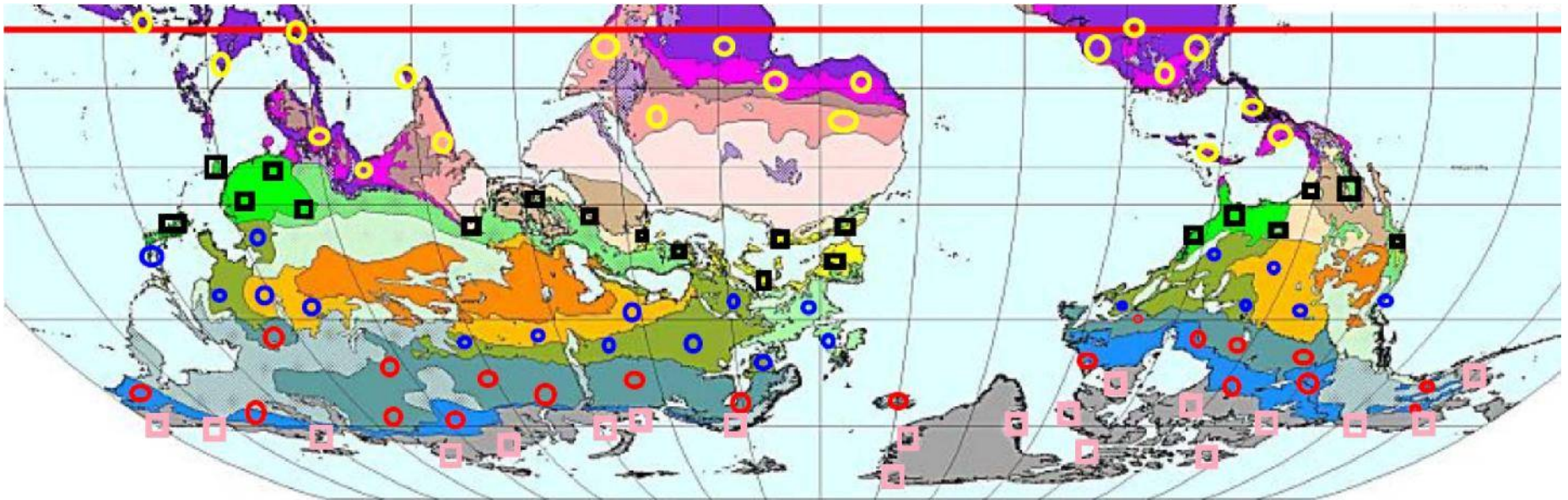


Duration of experiments:

Growth facility	Experimental duration (days)	n
Growth chamber	39	3100
Glasshouse	95	3500
Field	550	700



2. What causes the phenotypic differences? a. Field conditions



Global ecological zones:

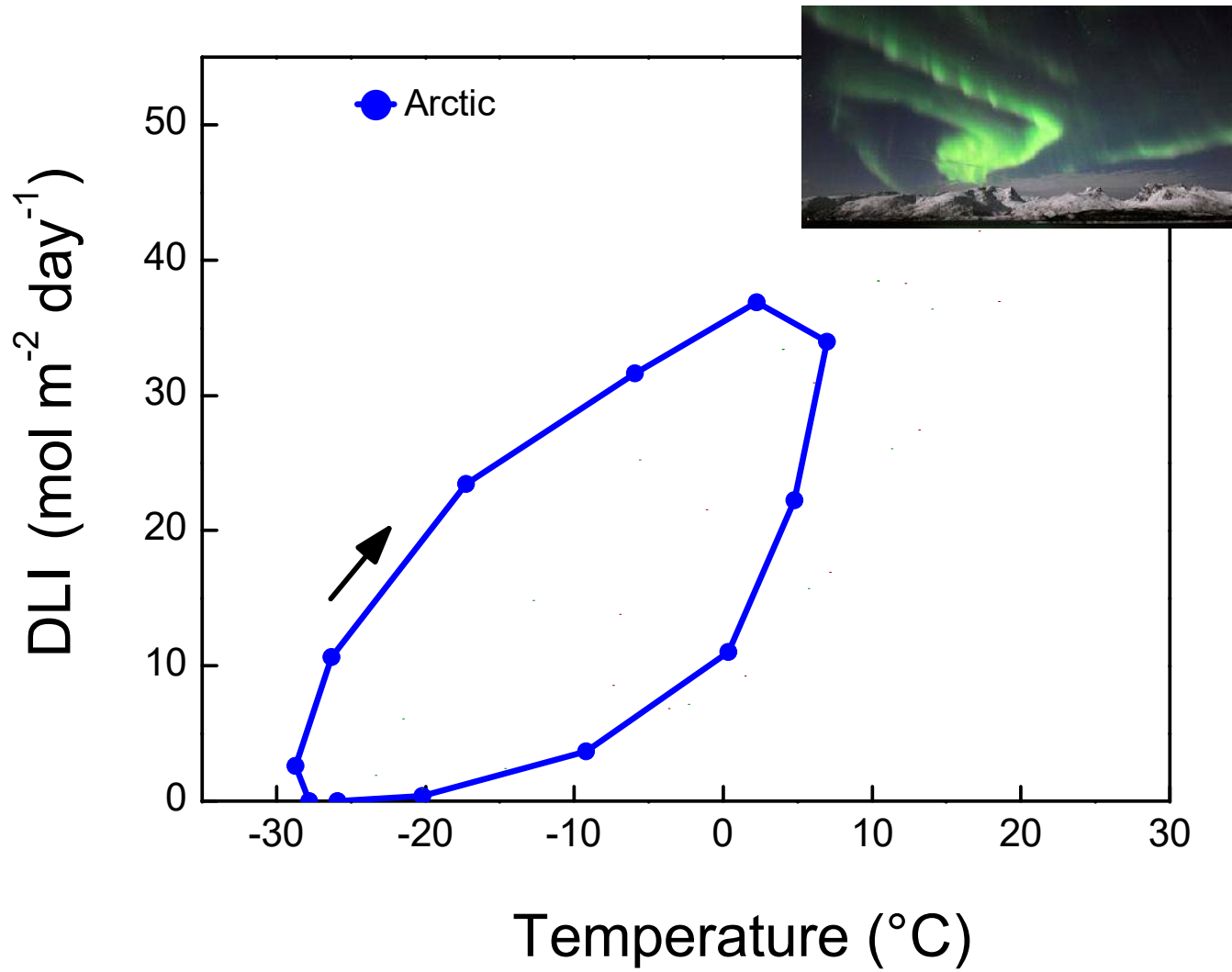
1. Arctic
2. Boreal
3. Temperate
4. Subtropical
5. Tropical

Excluded:

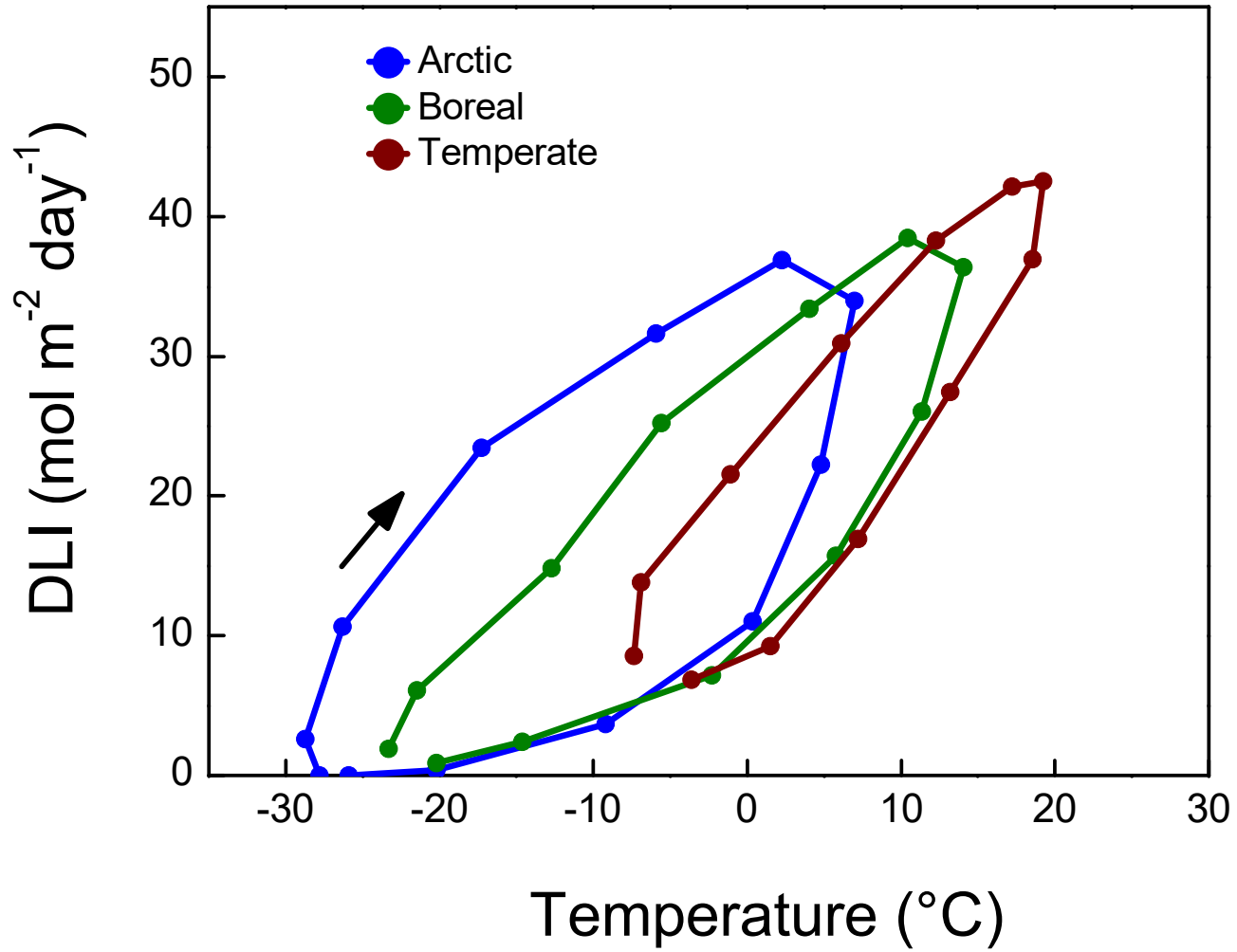
- Southern hemisphere
- mountains
- deserts

a. Field conditions:

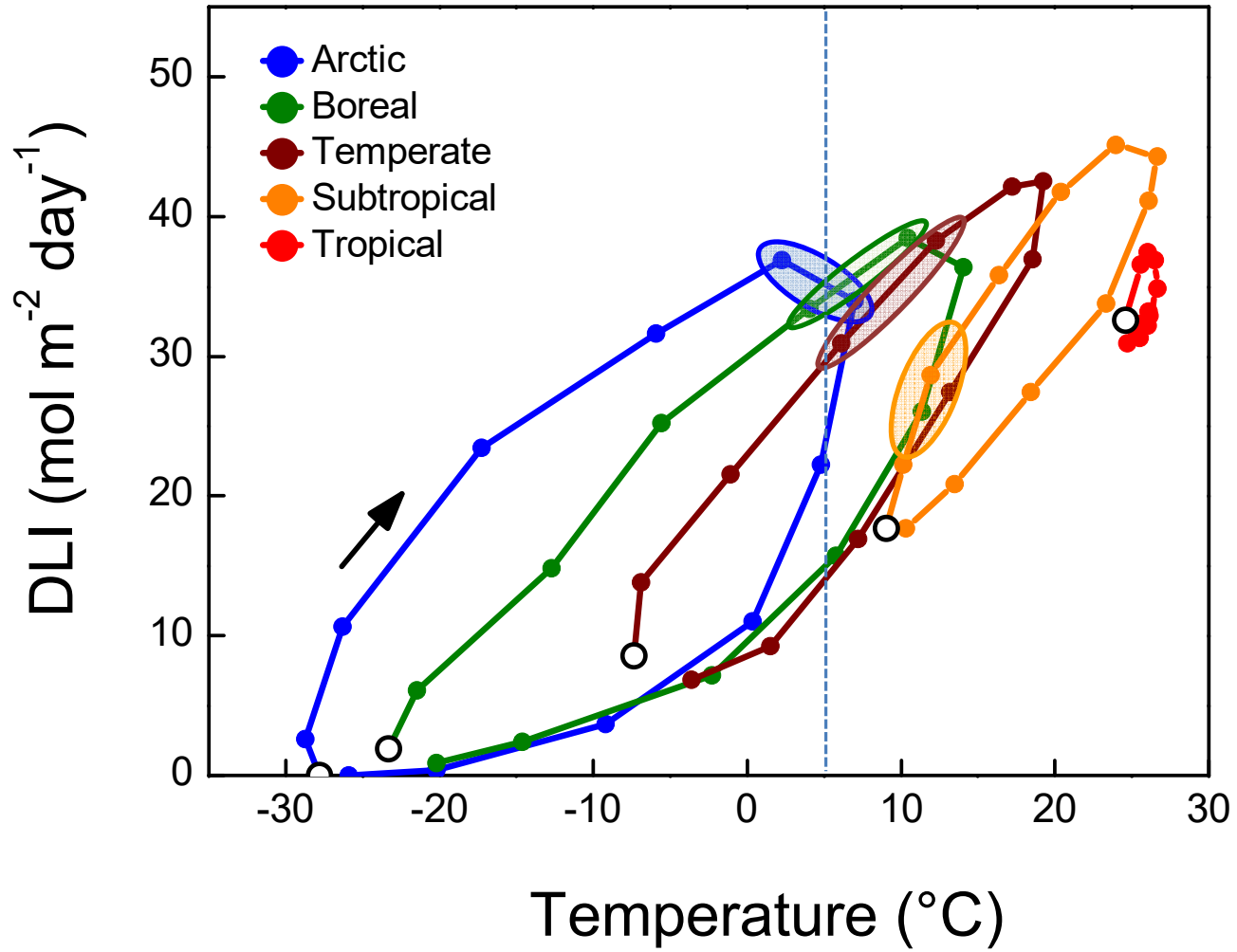
New et al. (1999)
Data for 1960-1990



b. Field conditions:



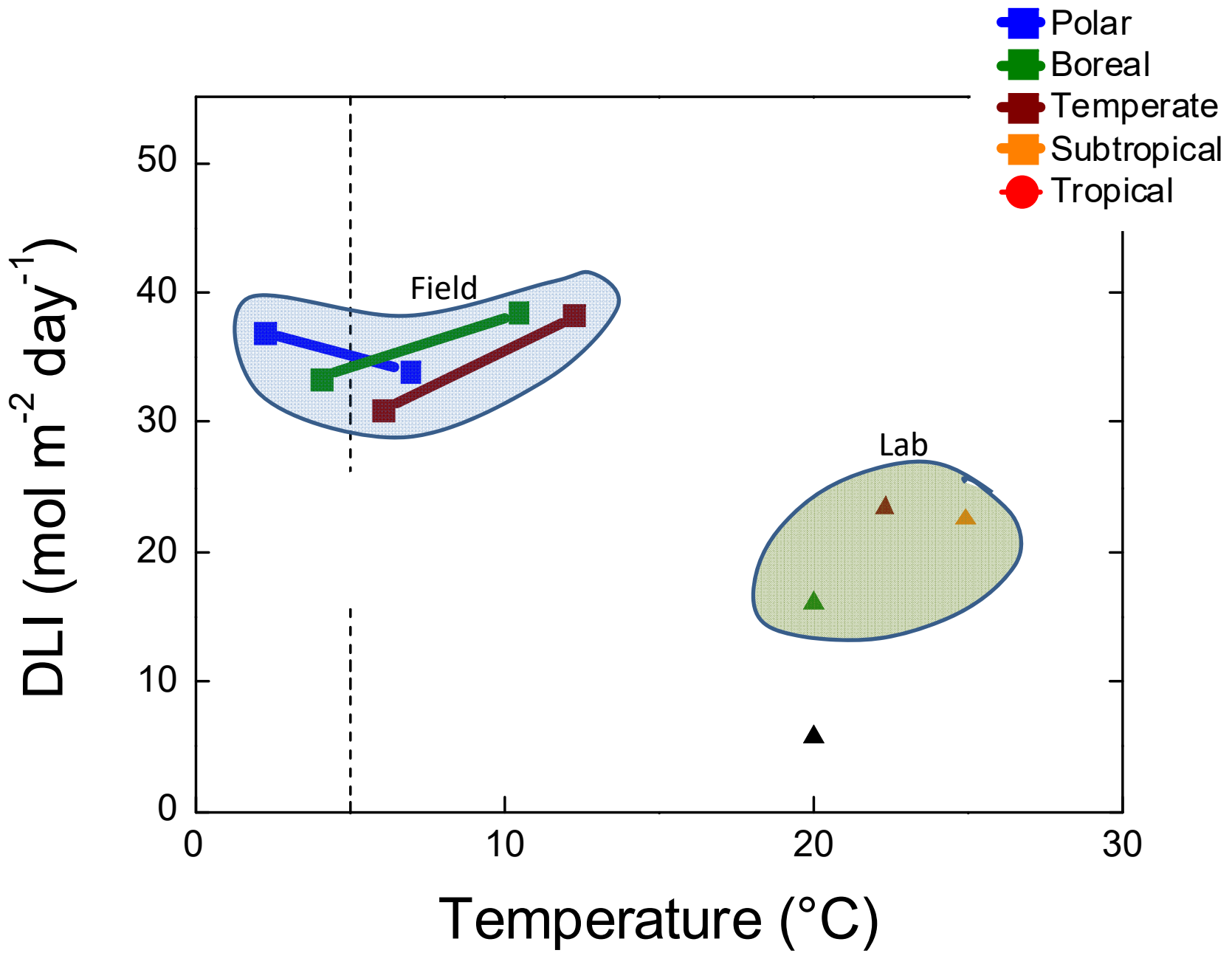
c. Field conditions:



2. Growth chamber conditions:



Species	Light (mol m ⁻² day ⁻¹)	Temperature (°C)
Arctic / Boreal	16	20
Temperate	23	22
(Sub)-tropical	23	25



Source- vs. sink-limitation:

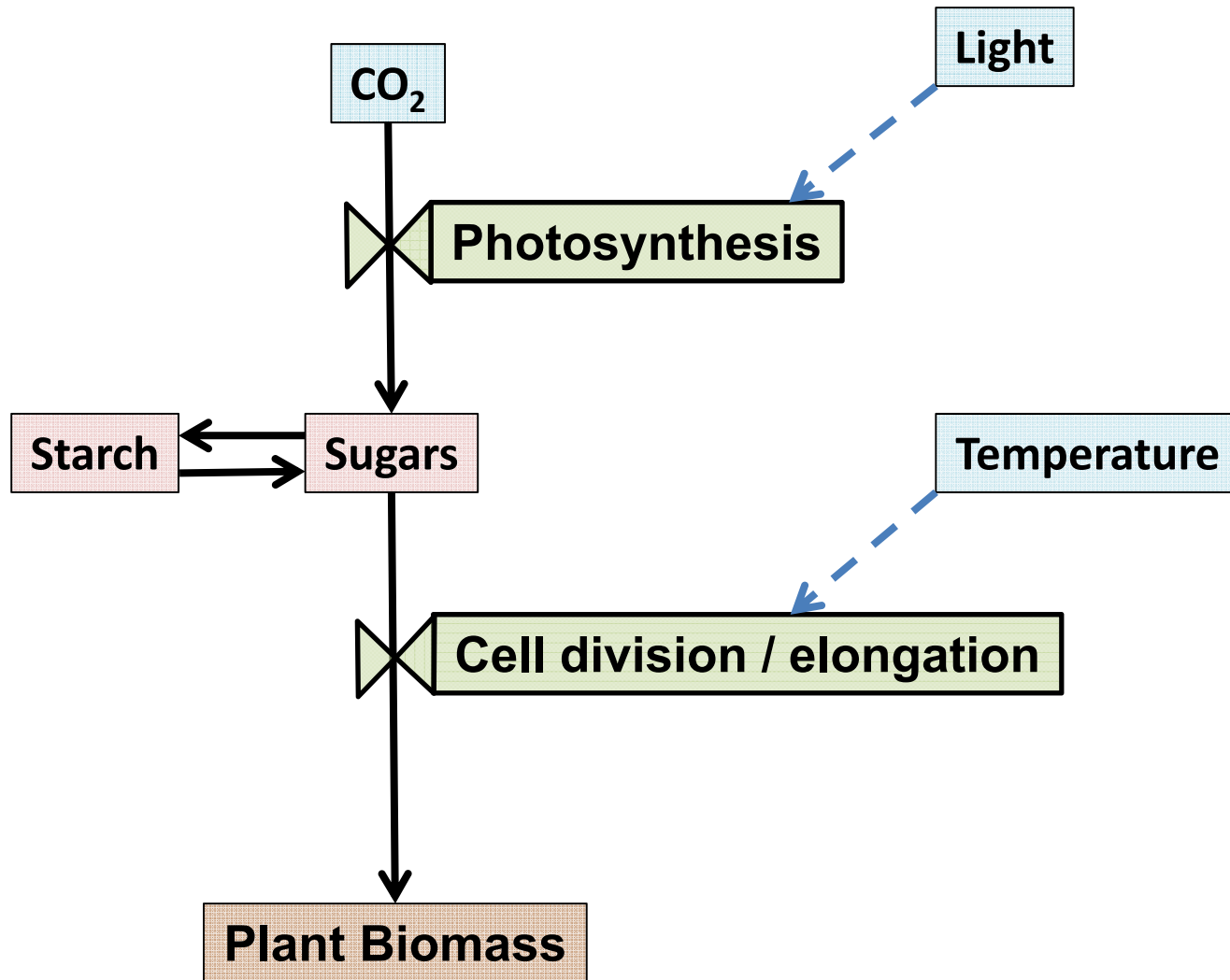


Photo-thermal ratio:

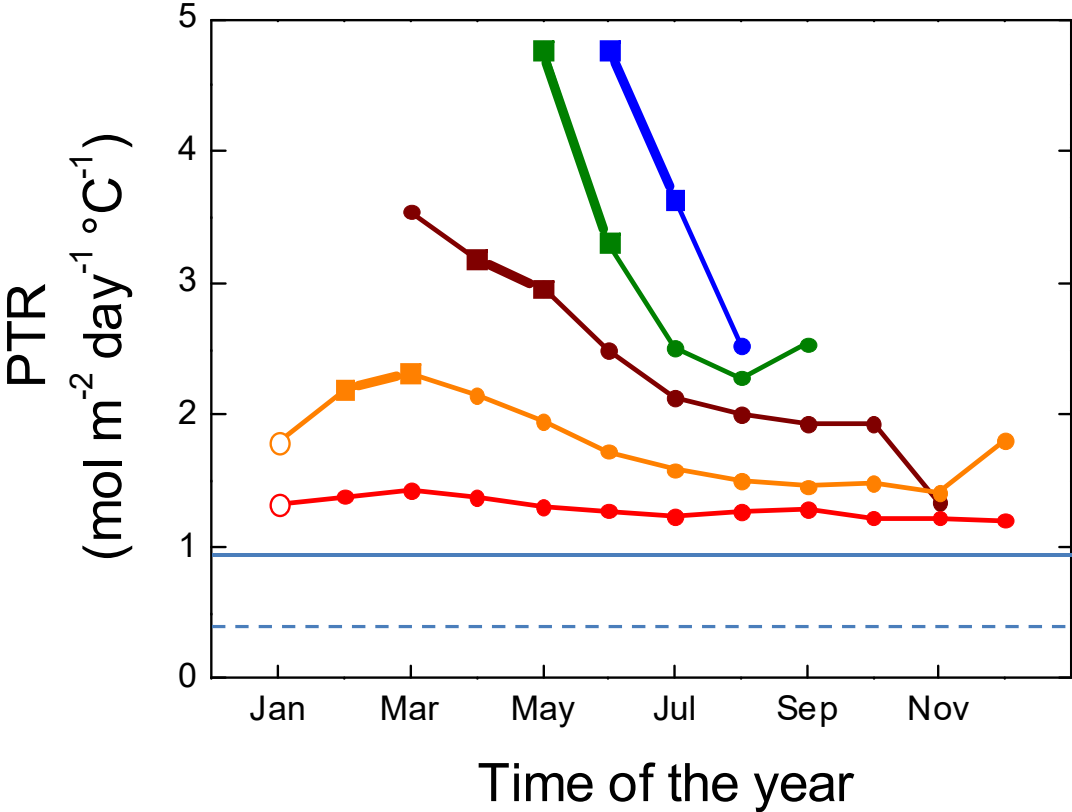
$$PTR = \frac{\overline{DLI}}{\overline{T}}$$



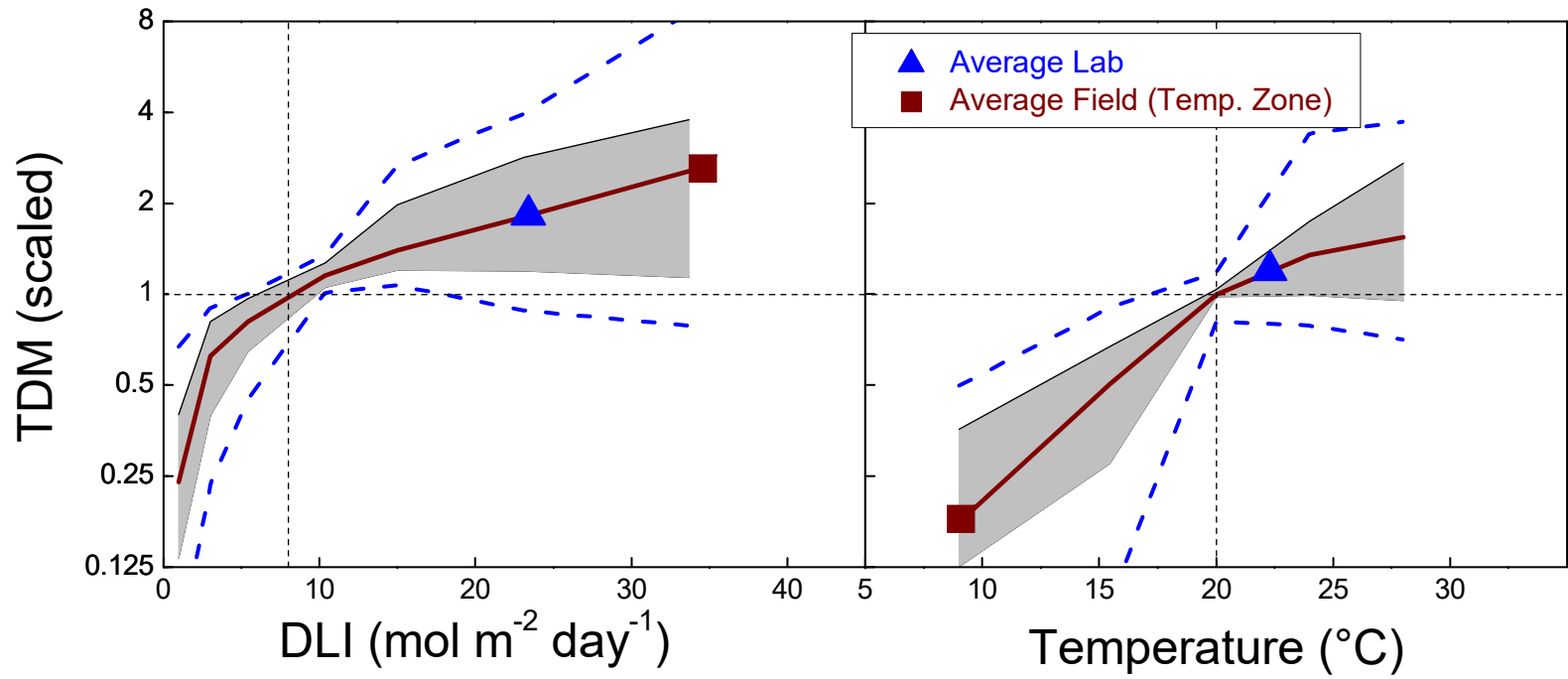
Fischer
(1985)



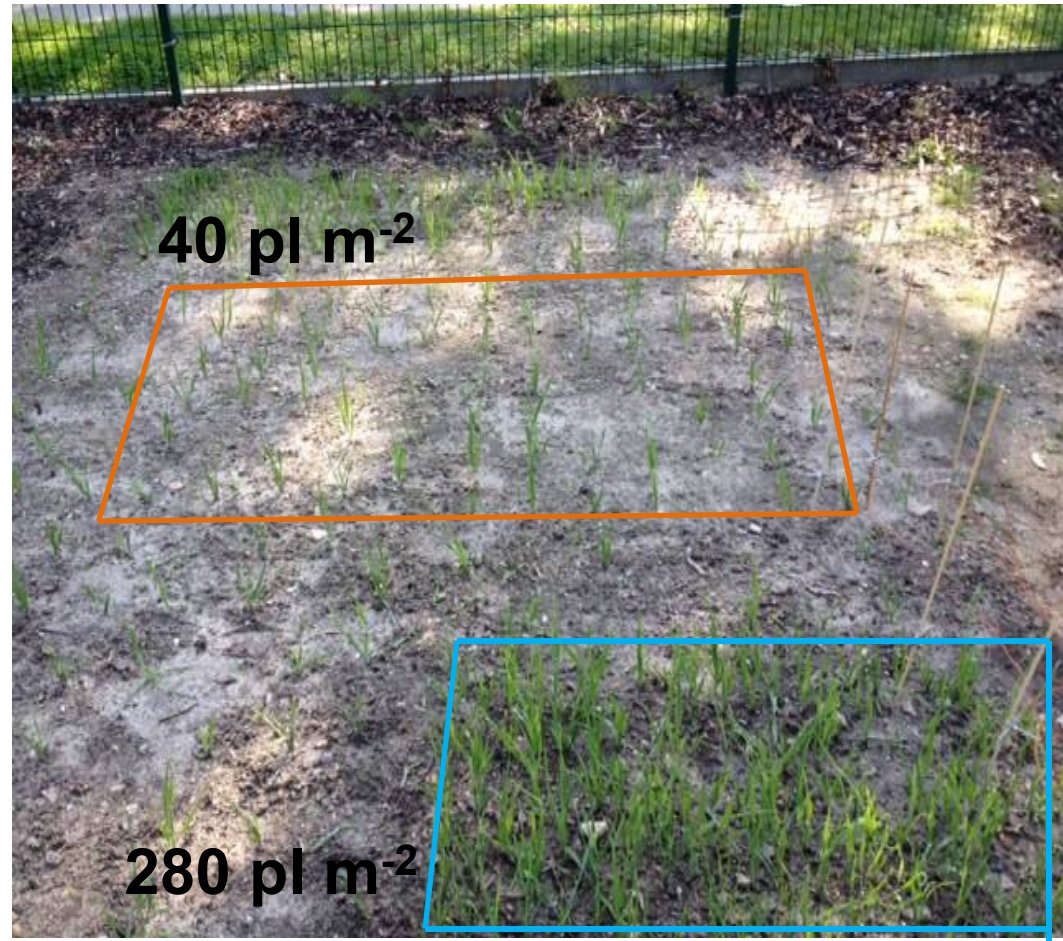
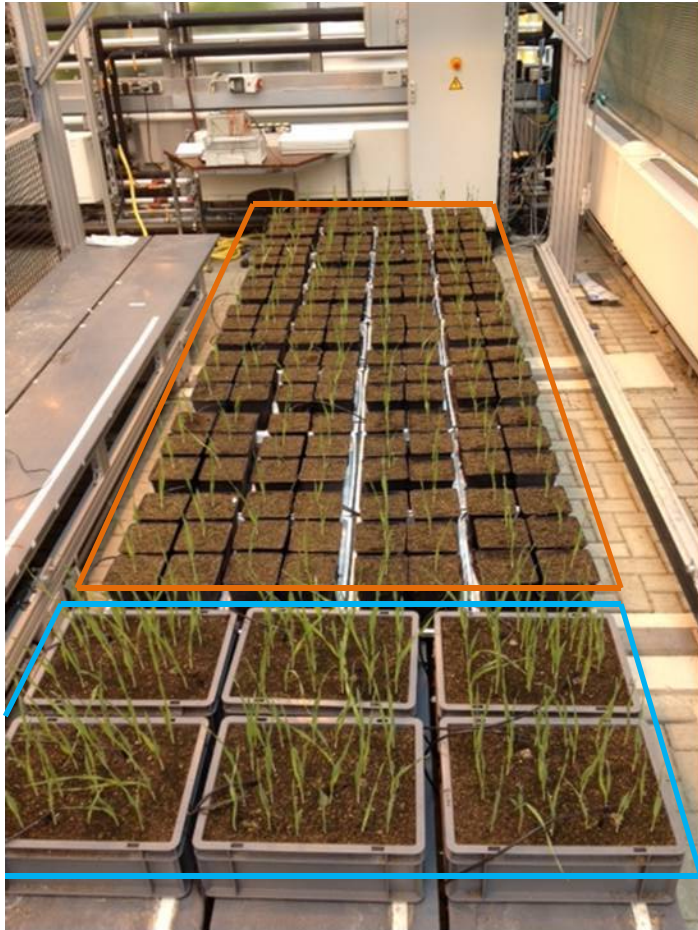
Heins
(1997)



Field vs. Lab:



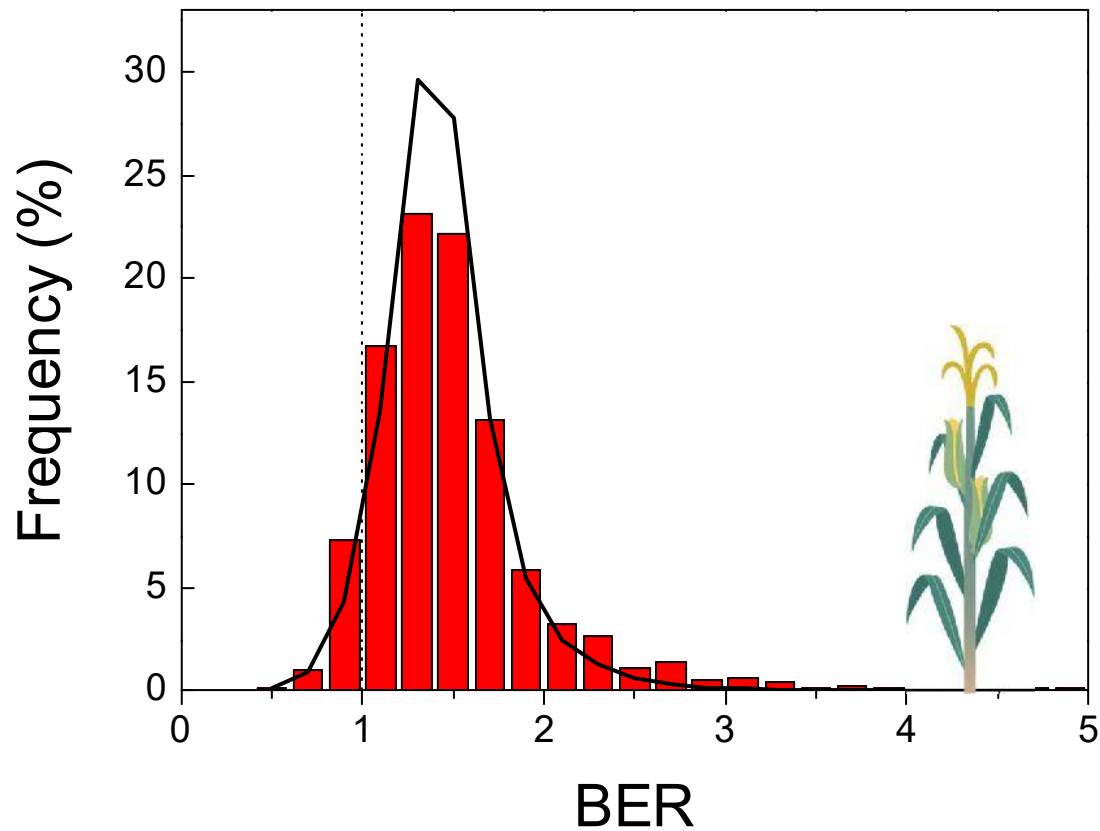
3. Plants in the field grow at higher densities than in the lab:



3. How useful is the analysis of individual plants? The case of elevated CO₂



Marie-Laure
Navas

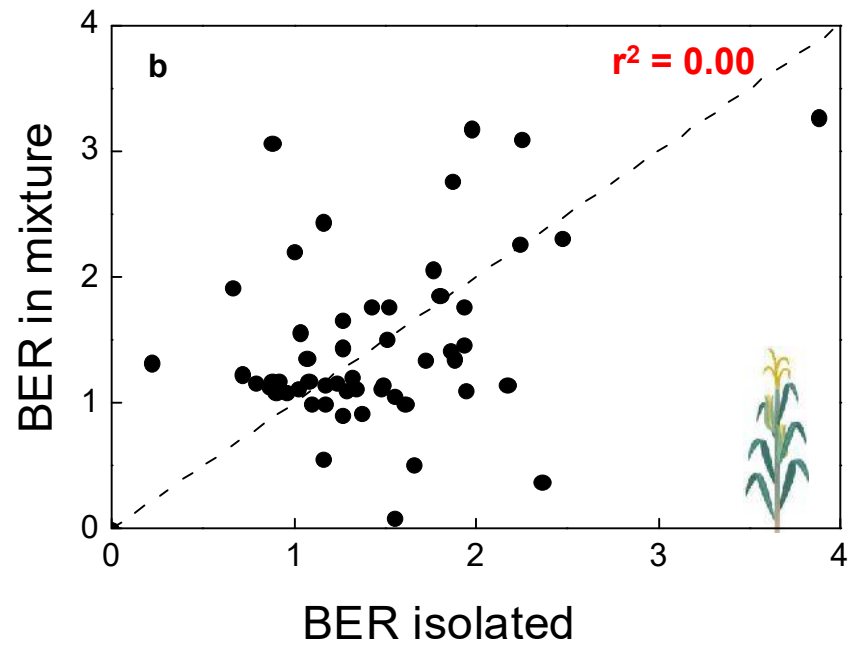


800 'experiments'
350 species

3. Can we now forecast the CO₂ response in a vegetation?



Marie-Laure Navas

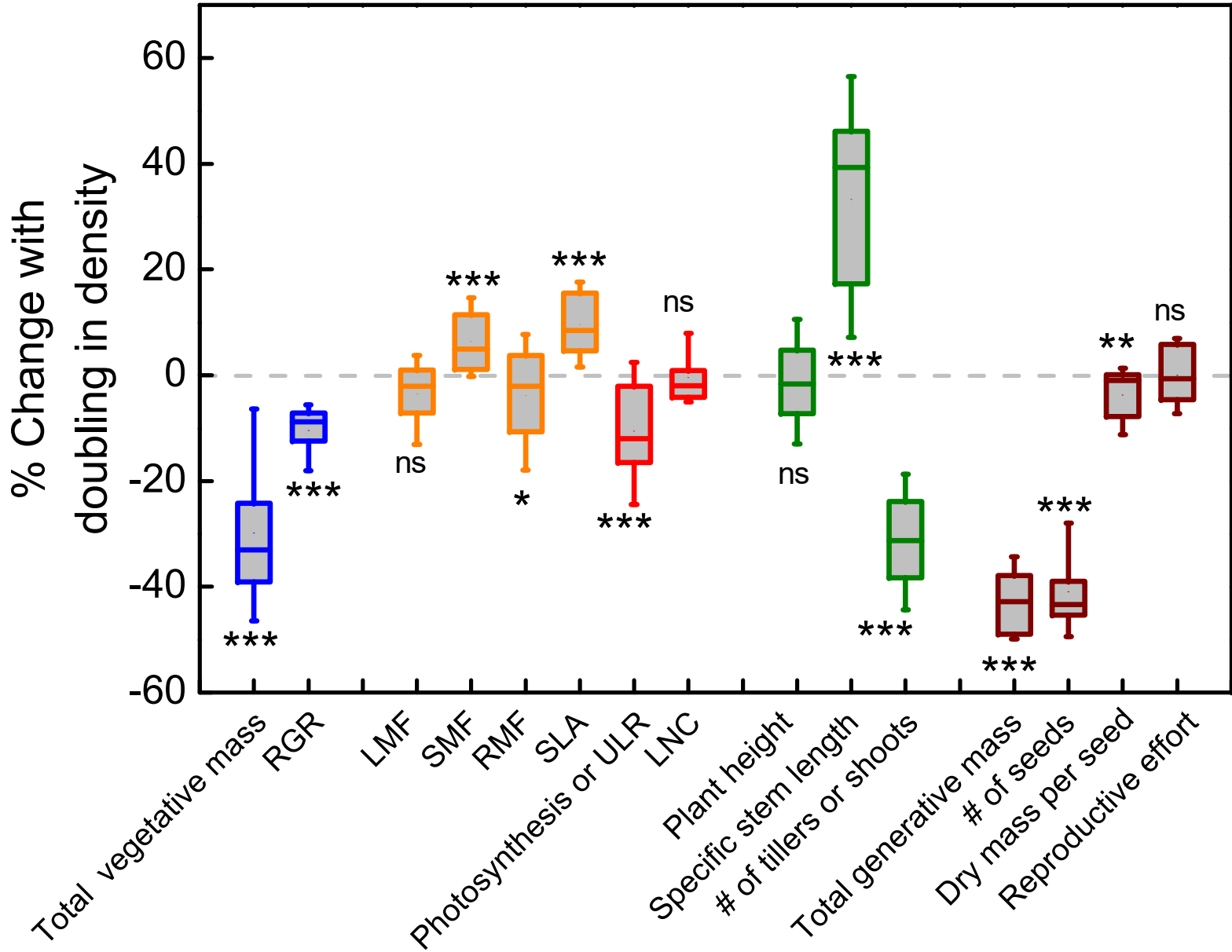


3. Plants in the field grow at higher densities than in the lab:

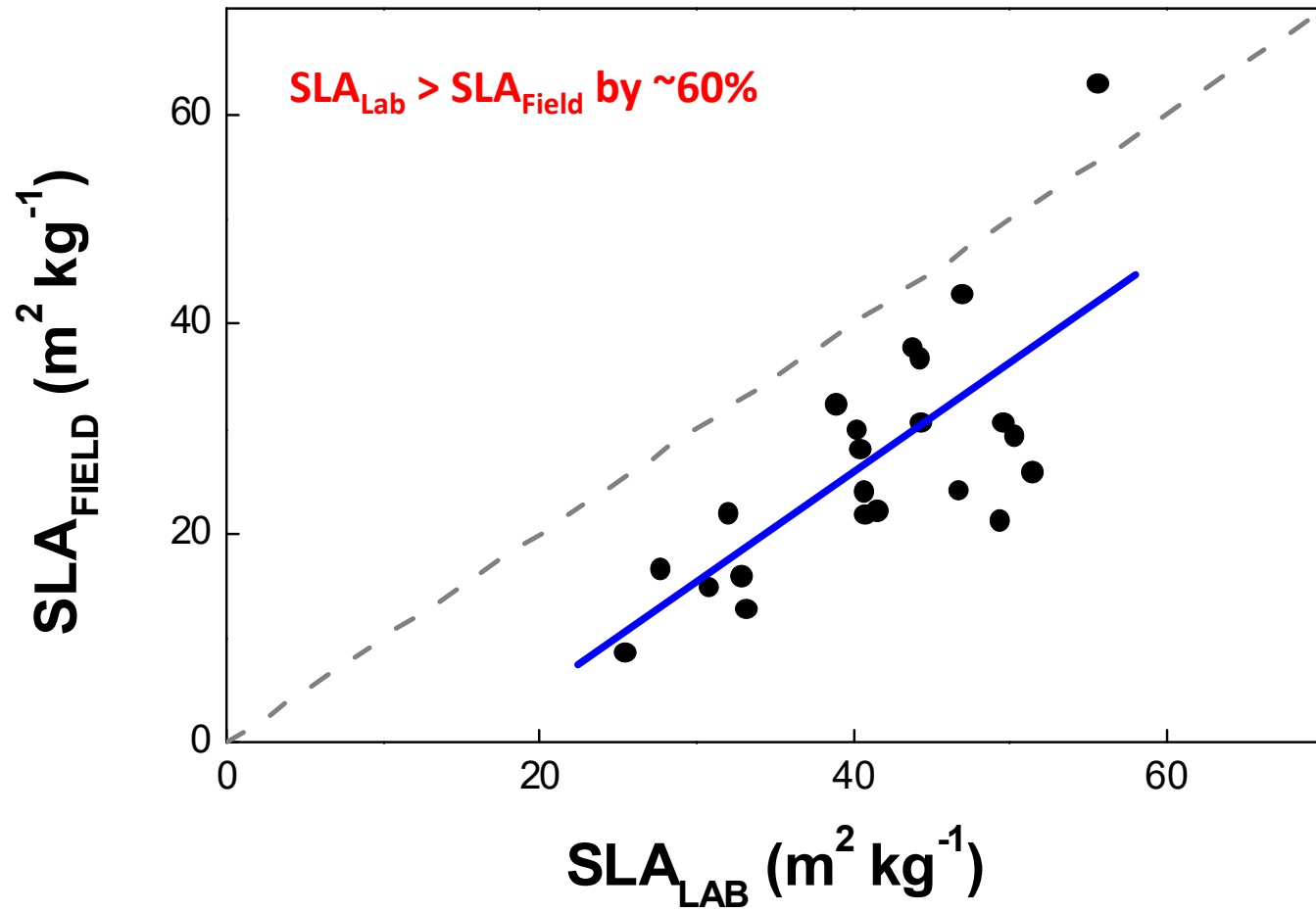


- Plants with high growth rates in the lab do not necessarily produce high crop yields in the field (Donald & Hamblin 1976; Cannell 1979).
- Modern *Zea mays* cultivars produce more because they perform better at high densities (Tollenaar & Wu 1998).

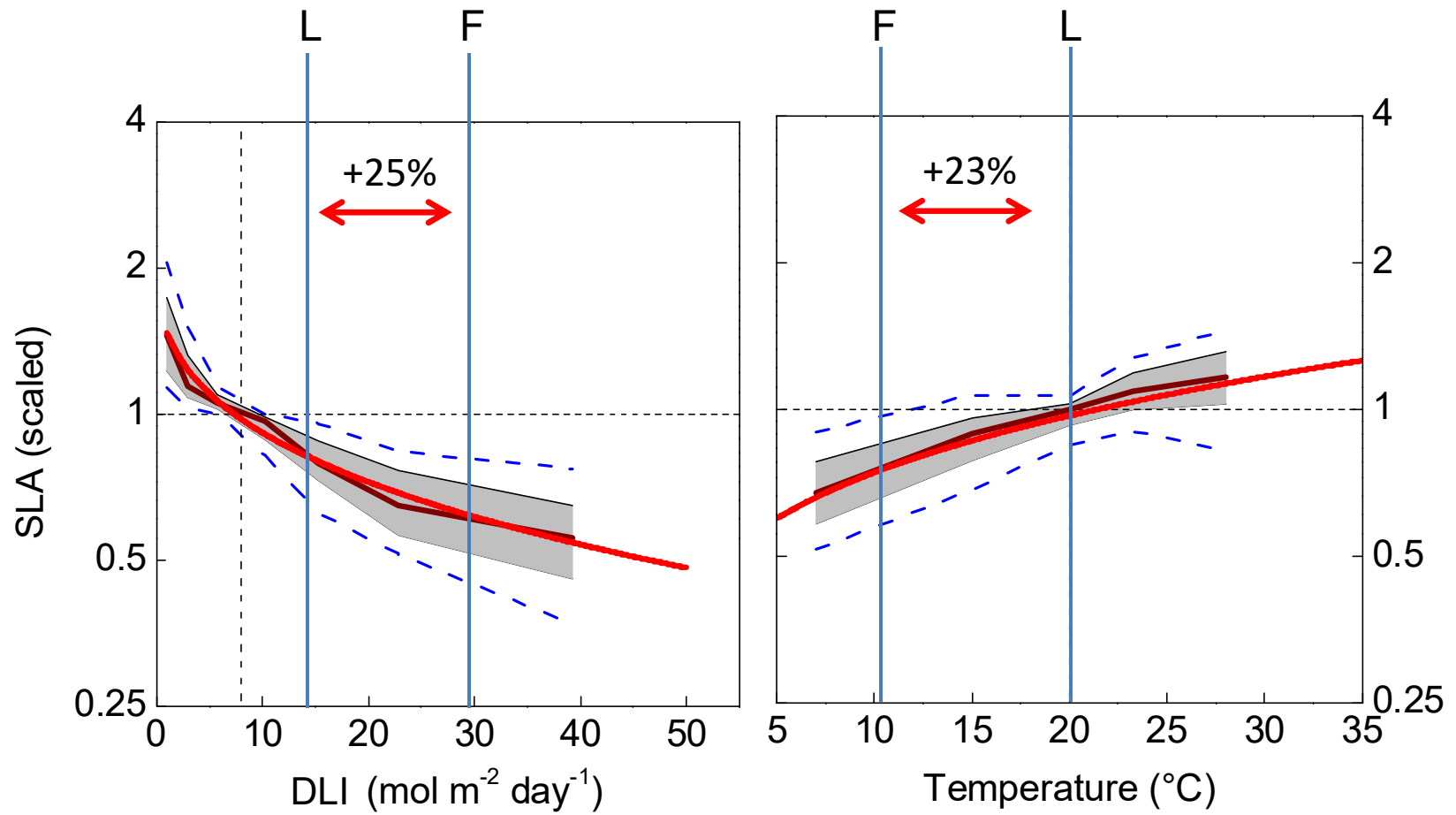
3. Effect of density:



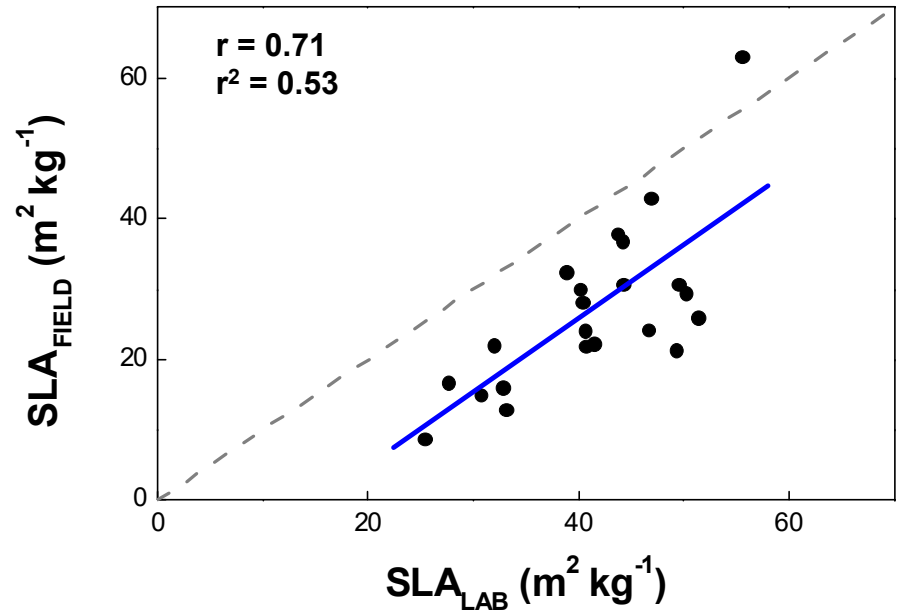
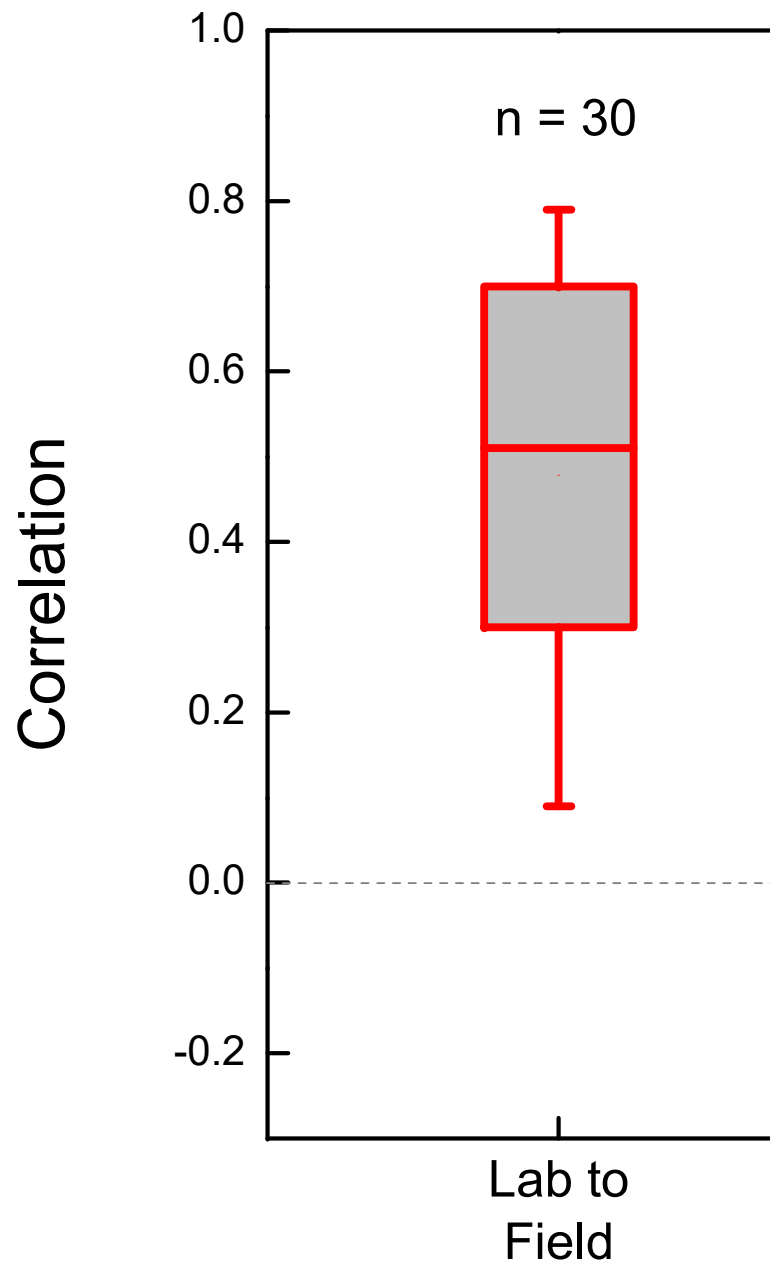
So can DPI and T explain the 60% difference between field and lab?



Can DPI and T explain the SLA difference between lab and field?

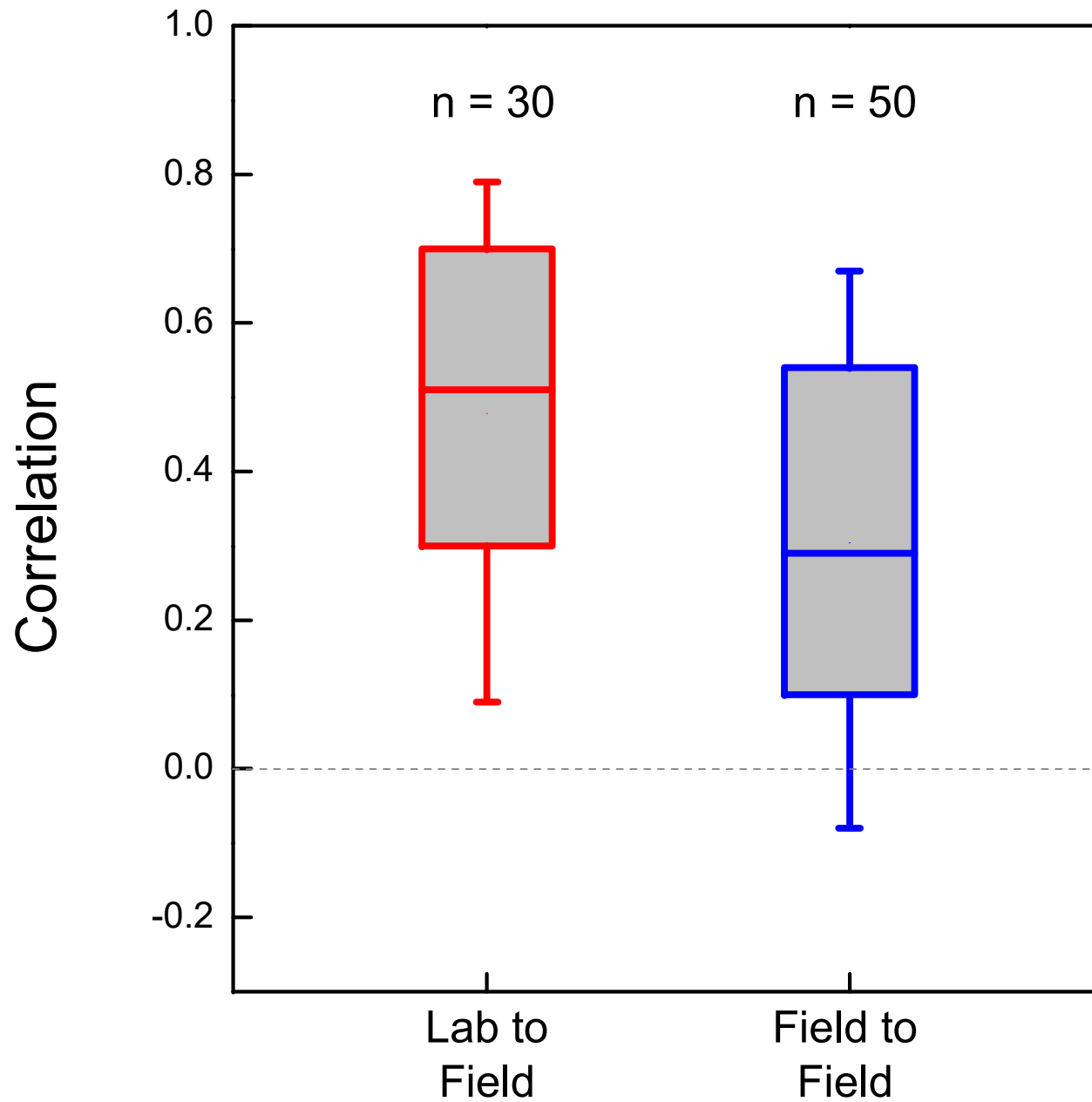


4. Correlation between lab and field results:



Median
 r^2
0.26

4. Correlation between lab and field results:



**Median
 r^2**

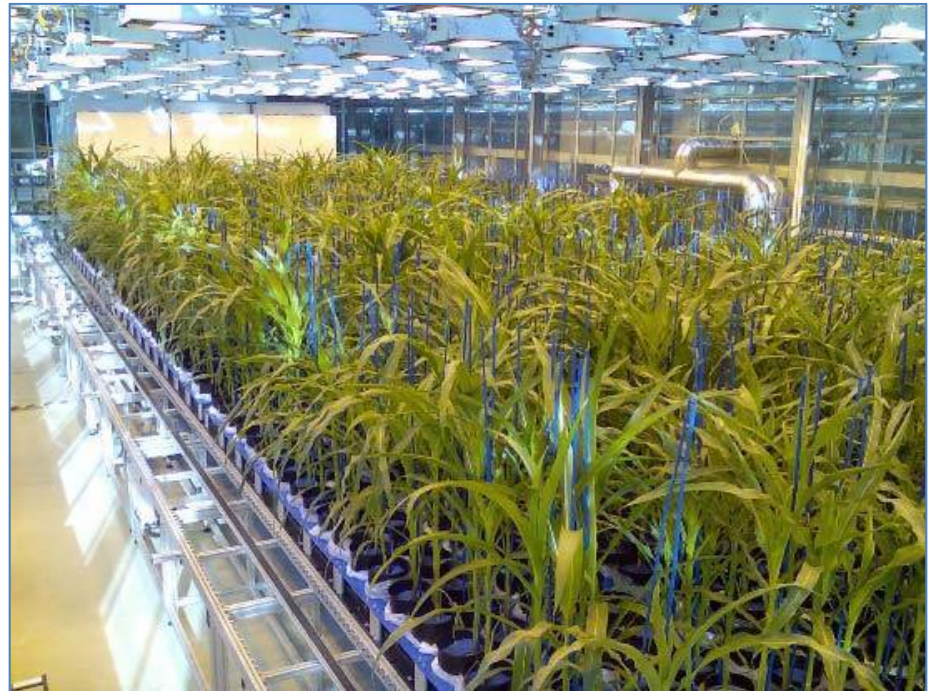
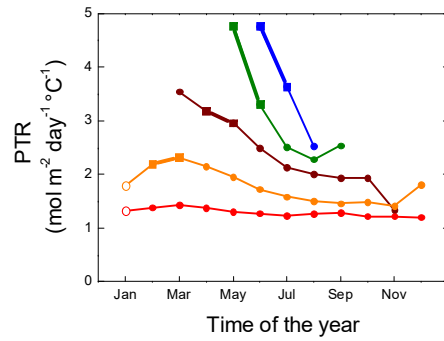
0.26

0.08

Improve the translation from lab to field:

Junker et al. (2015) FiPS

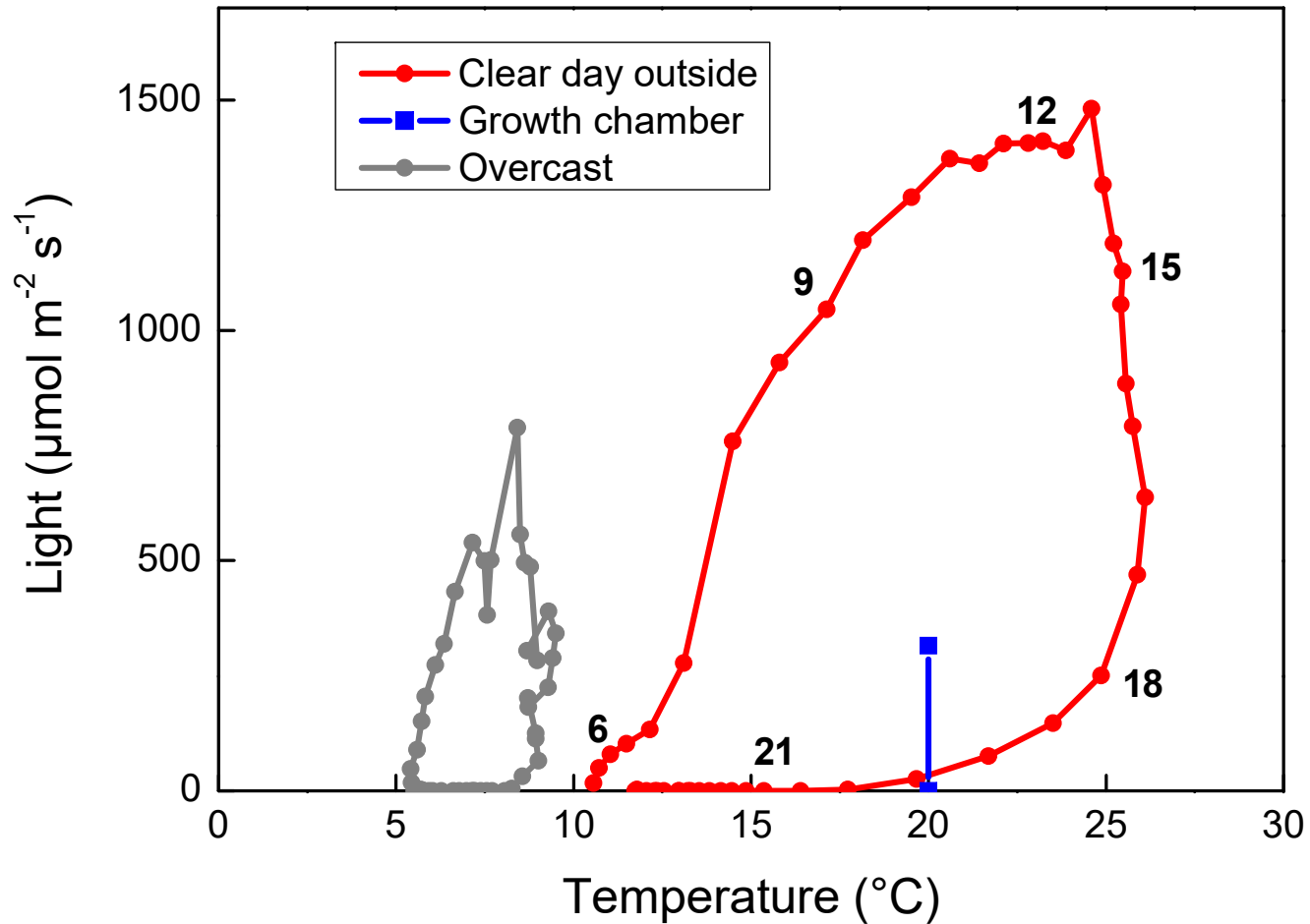
1. Use higher PTR



Lab-Field r^2
0.26 → 0.50

Improve the translation from lab to field:

3. Use conditions that fluctuate from day to day



How about applying the environmental conditions from outside?

Improve the translation from lab to field:

Hohmann et al. (2016) PCE

4: Grow plants at higher densities



(& big pots & foil house)

Lab - Field r^2
0.41 – 0.63

Other ways to improve the translation from lab to field:

presse.inra.fr
ciwr.ucanr.edu

5. Use stepping stones:

- Field soils
- Experimental gardens, OTC's, mesocosms



6. Lab and field: Better characterisation of soil and atmospheric conditions



7. Think in dose-response curves

8. Use simulation models



Conclusions:



1. In growth chambers:
 - Light levels are low
 - Temperature is high
 - strongly source limited
2. Responses of plants to environmental stresses are generally maintained
3. Correlations across species/genotypes are maintained to some extent

See also:

[Poorter et al. \(2016\)](#)

[Pampered inside, pestered outside? Differences and similarities between plants growing in controlled conditions and in the field. New Phytol. 212: 838-855](#)

Thanks to:



Johannes
Postma



Uli Schurr



Michael Kleyer



Tobias
Wojciechowski



Roland
Pieruschka



Wim van der
Putten



Fabio Fiorani